

# Supplementary information

## Materials and methods

### Culture conditions and chemical treatments

*G. lucidum* was cultured in seed culture as previously described (Cui et al., 2017), then inoculated (10%, volume fraction) into fermentation culture at 28 °C for 7 d with shaking at 180 r/min. The fermentation culture containing the following components were prepared: malt extract (20 g/L), yeast extract (18 g/L), KH<sub>2</sub>PO<sub>4</sub> (3 g/L), MgSO<sub>4</sub> (1.5 g/L), and vitamin B<sub>1</sub> (0.05 g/L) at an initial pH of 5.5.

All chemicals were obtained from Sigma (St Louis, MO, USA) and Solarbio (Beijing, China). ABA was added to the fermentation cultures on Day 4 of culture at concentrations of 0, 100, 200, 300, 400, and 500 µmol/L. After ABA treatment for 24 h, CaCl<sub>2</sub> (10 mmol/L), ethylene glycol bis (2-aminoethyl), tetraacetic acid (EGTA, 10 mmol/L), and LaCl<sub>3</sub> (10 mM) were added on Day 5, respectively, and incubated for 24 h for Ca<sup>2+</sup> signaling analysis. Mycelium was collected on Day 6 of culture.

### Measurement of total GT content

The mycelium was collected and mixed with 95% ethanol at a ratio of 1:50 (volume fraction) and extracted twice for 20 min at 50 °C under ultrasound (400 W). The vanillin-glacial acetic acid-perchloric acid method was used for GT content determination. Successively, 0.1 mL of extracts was evaporated solvent with heating at 60 °C, and then 0.2 mL of 5% vanillin-glacial acetic acid solution (mass fraction) and 0.5 mL of perchloric acid were added and incubated at 60°C for 20 min, then cooled to the ambient temperature. The GT content was measured at 550 nm after 5 mL of glacial acetic acid was added.

### Real-time quantitative polymerase chain reaction analysis

Total RNA preparation and RT-PCR were carried out as previously reported (Gu et al., 2017). The key genes of GT biosynthesis used were as follows: 3-hydroxy-3-methylglutaryl-CoA reductase (*hmgr*, accession number EU263989), squalene synthase (*sqs*, accession number DQ494674), and lanosterol synthase (*ls*, accession number FJ195872). The key genes involved in Ca<sup>2+</sup> signaling included calcium-channel protein *cch1*, calcium transporting adenosine triphosphatase (ATPase), calcium-dependent mitochondrial carrier protein (CDMCP), and vacuolar calcium ion transporter, calcium/calmodulin-dependent protein kinase *cmkB* (selected via transcriptome sequencing; Table S1, Fig. S4). Relative abundance was determined using *gpd* and *β-Tub* as internal standards. The related primers are listed in Table S2.

### Determination of cytosolic Ca<sup>2+</sup> concentration

The sample was crushed and homogenized, and 1.0g of sample was added in a 40mL sampling bottle, fixed volume to 10mL with deionized water, and then extracted by ultrasound (400 W) for 30 min. The extracted solution was passed through a 0.45-µm filter membrane and prepared to be tested, CaCl<sub>2</sub> was measured as a standard.

The analysis of Ca<sup>2+</sup> concentration was performed on an ion chromatograph (ECO-C, Metromhm, Switzerland). The sample was separated on a Metrosep C4-150 column (250 mm×4.6 mm). The mobile phase consisted of HNO<sub>3</sub> (1.0 mmol/L and dipicolinic acid (0.7 mmol/L), and the detected volume was 20 µL with an elution rate of 1.5 mL/min at a column temperature of 25 °C. A conductance detector was used for measurement.

The Ca<sup>2+</sup> concentration was expressed as follows:

$$\text{Ca}^{2+} \text{ concentration } (\mu\text{g/g dry weight}) = \frac{\text{detected concentration } (\mu\text{g/mL}) \times \text{fixed volume (mL)}}{\text{sample fresh weight (g)}} / \text{ratio of dry weight to fresh weight of mycelium}$$

(1).

### Determination of antioxidant properties

A total of nine antioxidant indexes were determined, including the contents of H<sub>2</sub>O<sub>2</sub>, malondialdehyde (MDA), and glutathione (GSH), superoxide dismutase (SOD) activity, glutathione peroxidase (GSH-P<sub>X</sub>) activity, catalase (CAT) activity, peroxidase (POD) activity, and ascorbate peroxidase (APX) activity. The ability to resist O<sub>2</sub><sup>-</sup> was also measured. All these determined methods followed the manufacturers' instructions in the detection kits purchased from Nanjing Jiancheng Bioengineering Institute.

### Statistical analysis

All experiments were carried out in at least triplicate, and the results were expressed as the mean±standard deviation (SD),  $n=3$ . Statistical analysis was performed using IBM SPSS statistics 20 and Duncan's multiple-range test ( $P<0.05$ ).

### References

- Cui ML, Yang HY, He GQ, 2017. Apoptosis induction of colorectal cancer cells HTL-9 in vitro by the transformed products of soybean isoflavones by *Ganoderma lucidum*. *J Zhejiang Univ-Sci B (Biomed & Biotechnol)*, 18(12):1101-1112.  
<https://doi.org/10.1631/jzus.B1700189>
- Gu L, Zhong X, Lian DH, et al., 2017. Triterpenoid biosynthesis and the transcriptional response elicited by nitric oxide in submerged fermenting *Ganoderma lucidum*. *Process Biochem*, 60:19-26.  
<https://doi.org/10.1016/j.procbio.2017.05.029>

## Supplementary Tables

Table S1 GO annotation of Ca<sup>2+</sup> signaling-related transcripts after ABA treatment ( $P < 0.05$ )

ID	log <sub>2</sub> (fold change)	P	Length	Swissprot	Non-Redundant protein sequence
TRINITY_DN1841_c0_g1	3.107	<0.001	5214	sp Q9UTN1 OAC1_SCHP O Mitochondrial oxaloacetate transport protein OS=Schizosaccharomyces pombe (strain 972 / ATCC 24843) OX=284812 GN=oac1 PE=3 SV=1	VWO99184.1 <b>Calcium dependent mitochondrial carrier protein</b> [ <i>Ganoderma boninense</i> ]
TRINITY_DN379_c0_g1	2.066	<0.001	9683	sp P13586 ATC1_YEAST Calcium-transporting ATPase 1 OS=Saccharomyces cerevisiae (strain ATCC 204508 / S288c) OX=559292 GN=PMR1 PE=1 SV=1	AFX00736.1 <b>calcium transporting ATPase</b> [ <i>Ganoderma lucidum</i> ]
TRINITY_DN1158_c0_g1	1.992	<0.001	2770	sp Q9Y899 KCC1B_EME ND <b>Calcium/calmodulin-dependent protein kinase cmkB</b> OS=Emericella nidulans OX=162425 GN=cmkB PE=1 SV=1	PIL28908.1 transporter [ <i>Ganoderma sinense</i> ZZ0214-1]
TRINITY_DN11101_c0_g1	3.525	<0.001	6710	sp O14234 CCH1_SCHPO <b>Calcium-channel protein cch1</b> OS=Schizosaccharomyces pombe (strain 972 / ATCC 24843) OX=284812 GN=cch1 PE=3 SV=1	PIL32546.1 transporter [ <i>Ganoderma sinense</i> ZZ0214-1]
TRINITY_DN13_c0_g1	1.665	<0.001	6726	sp O59768 VCX1_SCHPO <b>Vacuolar calcium ion transporter</b> OS=Schizosaccharomyces pombe (strain 972 / ATCC 24843) OX=284812 GN=vex1 PE=3 SV=1	PIL24302.1 hypothetical protein GSI_14055 [ <i>Ganoderma sinense</i> ZZ0214-1]
TRINITY_DN1869_c0_g1	-2.187	<0.001	1789	sp O59731 YHXB_SCHPO Uncharacterized J domain-	VWP00319.1 <b>Calcium/prot</b>

				containing protein C3E7.11c OS=Schizosaccharomyces pombe (strain 972 / ATCC 24843) OX=284812 GN=SPBC3E7.11c PE=3 SV=1	<b>on exchanger</b> [ <i>Ganoderma boninense</i> ]
TRINITY_DN1673_c0_g1	-2.202	<0.001	3842	sp P42839 VNX1_YEAST Low affinity vacuolar monovalent cation/H(+) antiporter OS= Saccharomyces cerevisiae (strain ATCC 204508 / S288c) OX=559292 GN=VNX1 PE=1 SV=1	AVR29896.1 <b>putative calcium- hydrogen exchanger 4</b> [ <i>Ganoderma lucidum</i> ]
TRINITY_DN1040_c0_g1	-3.806	<0.001	5241	sp P40977 PLC1_SCHPO 1-phosphatidylinositol 4,5- bisphosphate phosphodiesterase 1 OS=Schizosaccharomyces pombe (strain 972 / ATCC 24843) OX=284812 GN=plc1 PE=1 SV=1	AVM41526.1 <b>phospholipase C</b> [ <i>Ganoderma lucidum</i> ]
TRINITY_DN5301_c0_g1	-1.499	0.037	1845	sp Q10063 EHS1_SCHPO <b>Calcium influx-promoting protein <i>ehs1</i></b> OS=Schizosaccharomyces pombe (strain 972 / ATCC 24843) OX=284812 GN=ehs1 PE=3 SV=1	AVM41529.1 Mid [ <i>Ganoderma lucidum</i> ]

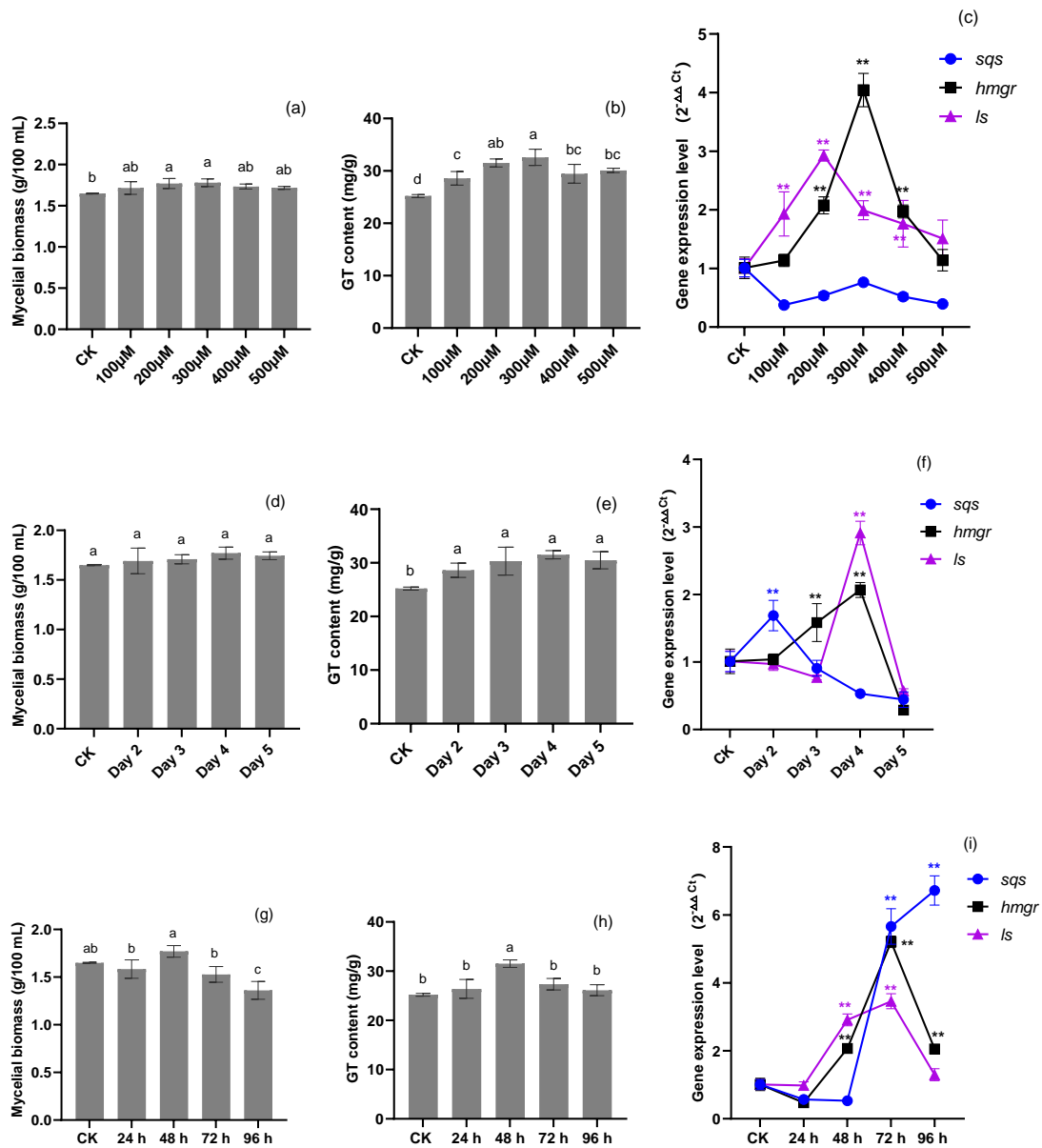
GO: Gene Ontology; ABA: abscisic acid; ID: identity; ATPase: adenosine triphosphatase.

**Table S2 Primer sequences used for reverse transcription-polymerase chain reaction analysis**

Gene ID	Primer name	Primer sequence (5' to 3')
<i>gpd</i>	Real-gpd-F	GATGAAGGACTGGCGTGGT
	Real-gpd-R	CCGTTGAGGCTGGGAATGAC
$\beta$ - <i>Tub</i>	Real- $\beta$ -Tub-F	GCGCTCTACGACATTTGCTT
	Real- $\beta$ -Tub-R	ACGATGGAGACGAGGTGGTT
<i>sqs</i>	Real-sqs-F	ACAGTTGTCAGCGAAGAGC
	Real-sqs-R	CGTAGTGGCAGTAGAGGTTG
<i>hmgr</i>	Real-hmgr-F	GTCATCCTCCTATGCCAAAC
	Real-hmgr-R	GGGCGTAGTCGTAGTCCTTC
<i>ls</i>	Real-ls-F	CTTCCGCAAGCACTACCCG
	Real-ls-R	AGCAGATGCCCCACGAGCC
calcium-channel protein <i>cch1</i>	Real-cch1-F	CCCTCCTCGTTCTCCTCCATCATAG
	Real-cch1-R	CATCGTCCTCAACTTCTGGCTCATC
calcium-dependent mitochondrial carrier protein	Real-CDMCP-F	CTCCTCCTGTCCCCTCTCTCAAC
	Real-CDMCP-R	GCTTGCTGTCCAGTCTGTCTAACC
calcium transporting ATPase	Real-calcium transporting ATPase-F	ATTCGTCCTGTCCGCATTCGTAAC
	Real-calcium transporting ATPase-R	TCGTCCCAATTCTCGAAACCTG
Vacuolar calcium ion transporter	Real-vacuolar calcium ion transporter-F	GAAGTCGGCGAAGAACAGAGTGAG
	Real-vacuolar calcium ion transporter-R	TGGATGGCGATGAAGAACAAGATGG
calcium/calmodulin-dependent protein kinase <i>cmkB</i>	Real-cmkB-F	CCCCGGAAGCATTGGTAAGT
	Real-cmkB-R	TGGGGCTCCCTGATTCCATA

ID: identity; F: forward; R: reverse.

## Supplementary figures



**Fig. S1** Effects of different ABA concentrations (0, 100, 200, 300, 400, and 500 μmol/L) (a-c), different ABA addition time points (Day 2, 3, 4, and 5 of cultivation) (d-f), and ABA treatment durations (24, 48, 72, and 96 h) (g-i) on the biomass, GT accumulation and gene expression level of *G. lucidum*. Data were expressed as the mean±standard deviation (SD),  $n=3$ . Different lowercase letters indicate significant differences among various treatments ( $P<0.05$ ).  $**P<0.01$ . ABA: abscisic acid; CK: contrast check; GT: ganoderic triterpenoids; *hmgr*: 3-hydroxy-3-methylglutaryl-CoA reductase; *ls*: lanosterol synthase; *sqs*: squalene synthase.

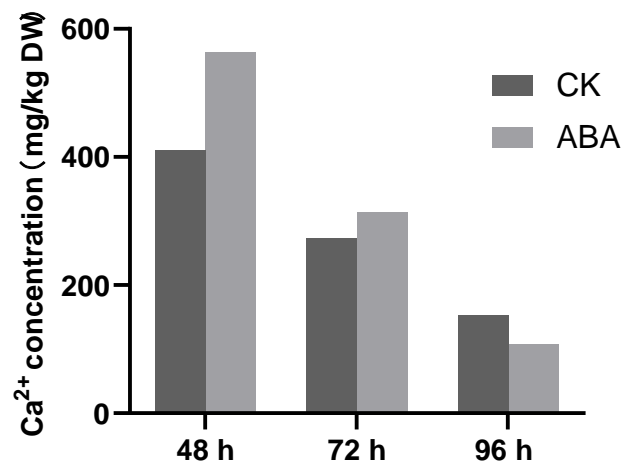
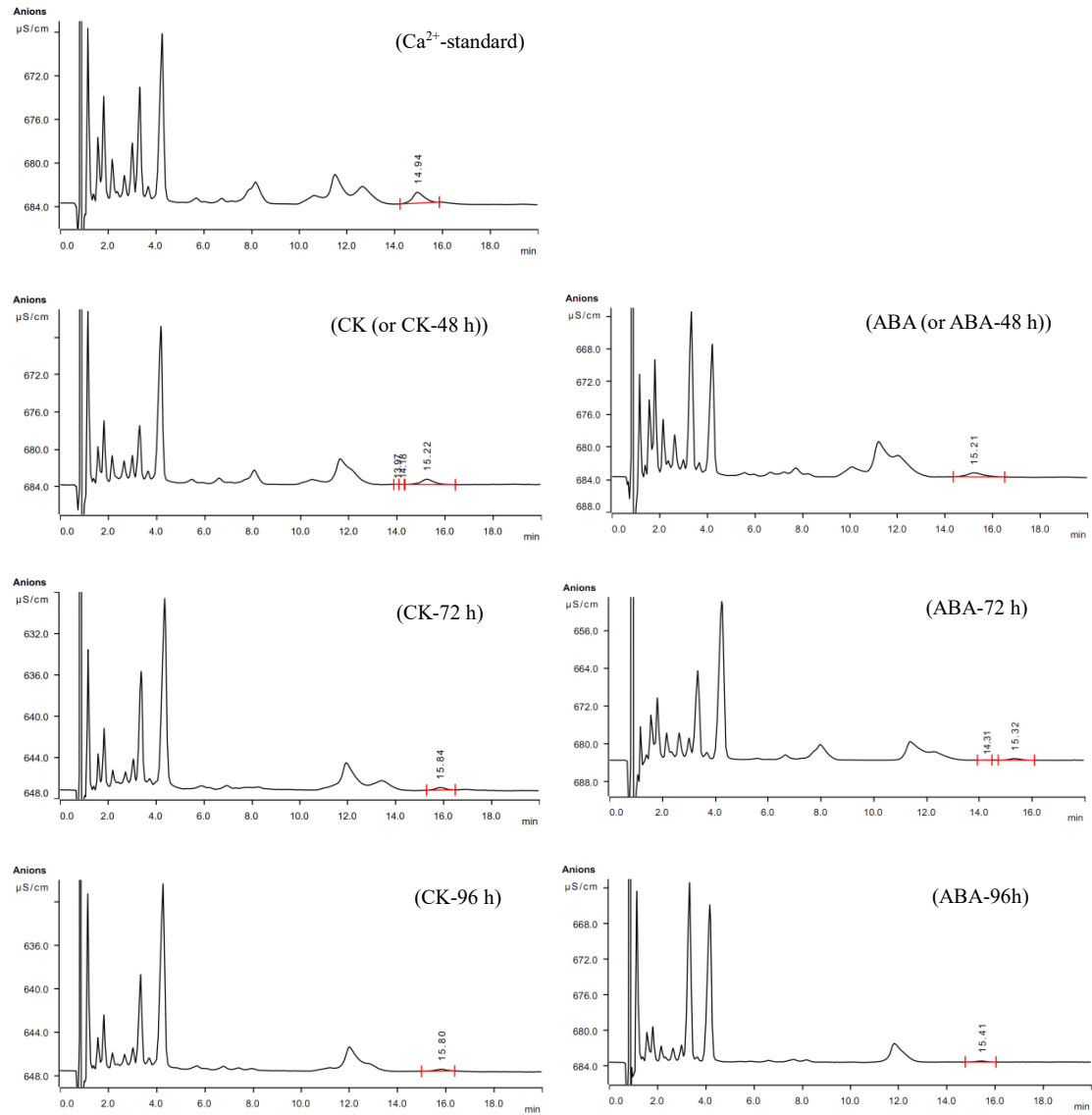


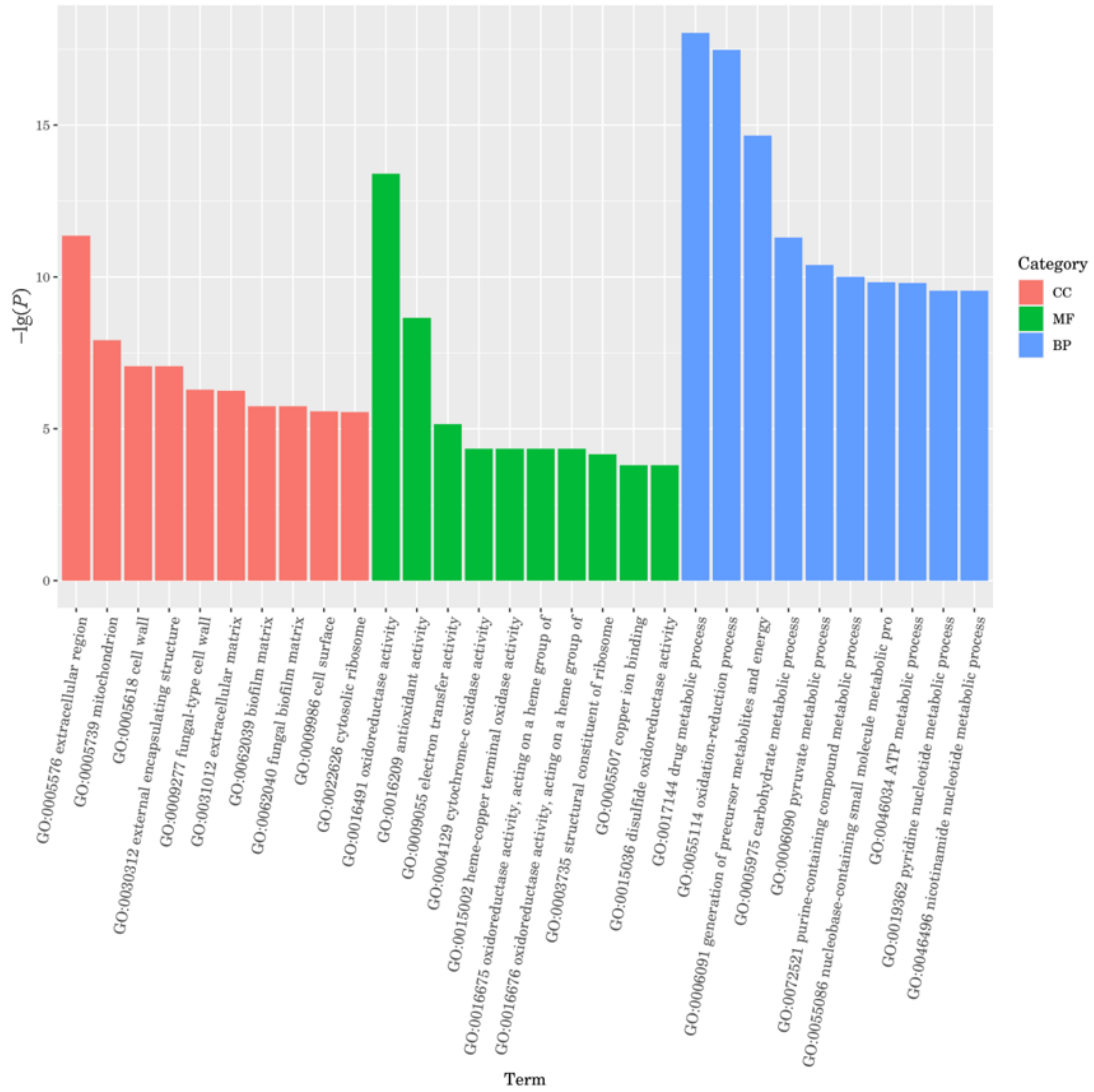
Fig. S2 Changes of cytosolic Ca<sup>2+</sup> concentration determining by ion chromatograph with ABA treatment for 48 to 96 h of *G. lucidum*. ABA: abscisic acid; CK: contrast check; DW: dry weight.



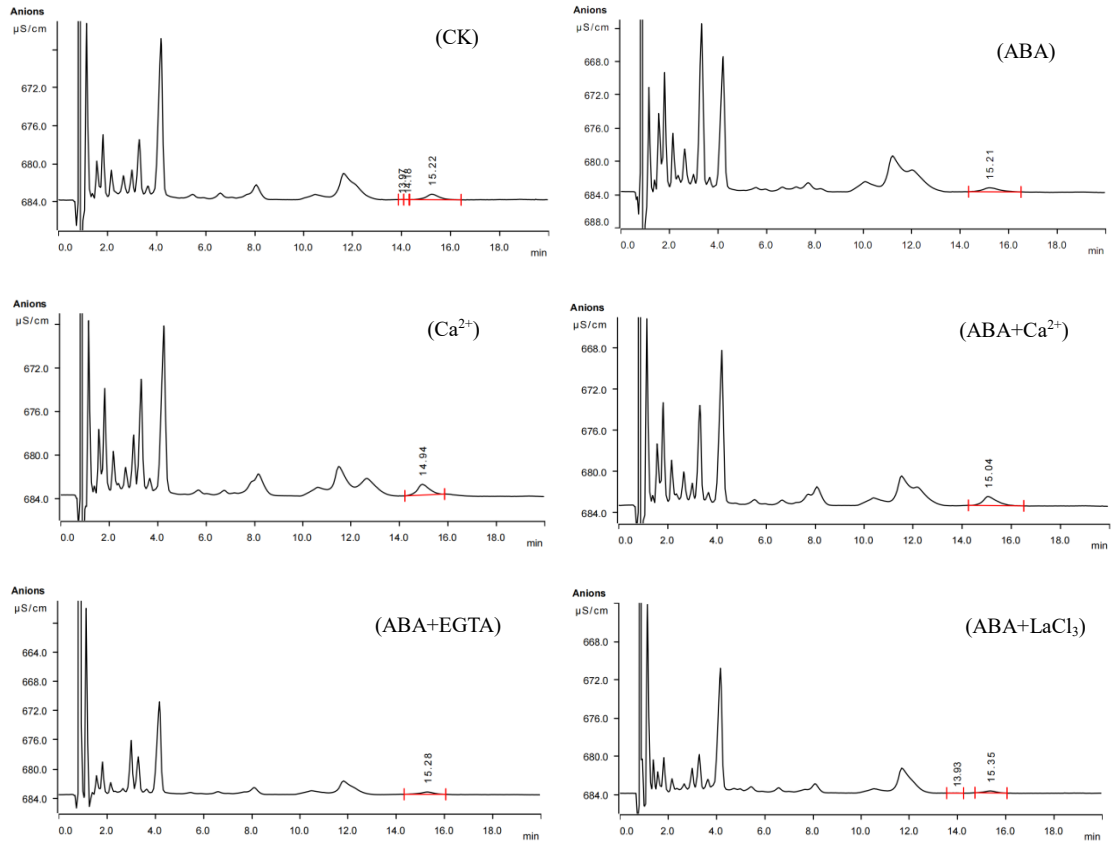
Sample	Retention time (min)	Ca <sup>2+</sup> concentration (mg/kg wet weight)	Ratio of dry weight to wet weight of mycelium	Ca <sup>2+</sup> concentration (mg/kg dry weight)
Ca <sup>2+</sup> -standard	14.767	7.682		
CK	15.218	39.742	0.097	409.711
CK-72 h	15.838	13.968	0.051	273.160
CK-96 h	15.798	10.197	0.067	153.051
ABA	15.208	46.706	0.083	563.485
ABA-72 h	15.315	19.916	0.063	313.677
ABA-96 h	15.412	7.015	0.065	108.061

**Fig. S3** Determination of cytosolic Ca<sup>2+</sup> concentration by ion chromatograph with ABA treatment for 48 to 96 h of *G. lucidum*. ABA: abscisic acid; CK: contrast check.





**Fig. S4** GO categories of differentially expressed unigenes identified after ABA treatment relative to control. Gene classification based on GO annotation,  $-\lg(P)$  was plotted on the Y-axis and the names of clusters were plotted on X-axis. GO: Gene Ontology; CC: cell component; MF: molecular function; BP: biological process.



Sample	Retention time (min)	Ca <sup>2+</sup> concentration (mg/kg wet weight)	Ratio of dry weight to wet weight of mycelium	Ca <sup>2+</sup> concentration (mg/kg dry weight)
CK	15.218	39.742	0.097	409.711
ABA	15.208	46.706	0.083	563.485
Ca <sup>2+</sup>	14.935	66.768	0.063	1062.946
ABA+Ca <sup>2+</sup>	15.042	69.025	0.056	1227.111
ABA+EGTA	15.283	24.128	0.048	505.998
ABA+LaCl <sub>3</sub>	15.352	13.074	0.145	90.428

**Fig. S5 Determination of cytosolic Ca<sup>2+</sup> concentration by ion chromatograph with different treatments mediated by ABA. ABA: abscisic acid; CK: contrast check; EGTA: ethylene glycol tetraacetic acid.**