

## Representative appressorium stage cDNA library of *Magnaporthe grisea*\*

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**Abstract:** A mature appressorium cDNA library of rice blast fungus, *Magnaporthe grisea*, was constructed in a  $\lambda$ TriplEx2 vector by SMART<sup>TM</sup> cDNA library containing  $2.37 \times 10^6$  independent clones about 100% of which harbor foreign cDNA inserts with average size of 660 bp. Of 9 randomly selected clones, 2 expressed sequence tags (ESTs) sequences did not have homologous EST sequences of *M. grisea* in GenBank. The appressorium cDNA library is suitable for gene expression analysis and function analysis of the late stages of appressorium formation and the early stages of penetration of *M. grisea*.

**Key words:** *Magnaporthe grisea*, Appressorium, cDNA library

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### INTRODUCTION

The rice blast fungus *Magnaporthe grisea* causes one of the most destructive diseases of rice. Genetic studies of this important pathogen during the past decade made it excellent for investigating fungal-plant interactions. Appressorium is an infection structure of *M. grisea*. Many genes expressed in appressorium, such as *PMK1* (Xu and Hamer, 1996), *CYP1* (Viaud *et al.*, 2002), *MAGB* and *MAC1* (Choi and Dean, 1997; Liu and Dean, 1997), *GAS1* and *GAS2* (Xue *et al.*, 2002), and *MPG1* (Lau and Hamer, 1996; Talbot *et al.*, 1993; 1996), are required for appressorium formation and pathogenesis by the fungus. Many approaches, such as expressed sequence tags (ESTs) sequencing (Kamakura *et al.*, 1999; Kim *et al.*, 2001; Rauyaree *et al.*, 2001), SAGE (Serial analysis of gene expression) (Irie *et al.*, 2003), microarray (Takano *et al.*, 2003), and mutants (Balhadere *et al.*, 1999; Shi *et al.*, 1995; Sweigard *et al.*,

1998), have been used to identify genes already expressed in appressorium. By now, more than 10000 ESTs have been sequenced from several cDNA libraries representing different stages of fungal growth, differentiation and plant infection of the pathogen, including appressoria, conidia, and mycelia from complete and nitrogen starvation cultures (Xu and Xue, 2002). However, appressorium cDNA libraries in existence were only constructed from conidia germinated for less than 8 h on an inductive surface (Kamakura *et al.*, 1999; Takano *et al.*, 2003), causing a significant problem for completely evaluating gene expression during the late stages of appressorium formation. We report here the result of producing an appressorium stage cDNA library of *M. grisea* from mature appressoria formed for 23.5–24.5 h.

### MATERIALS AND METHODS

#### *M. grisea* isolate, appressorium induction and RNA isolation

The appressorium of *M. grisea* GUY11 was used to isolate total RNA for construction of cDNA library.

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For appressorium formation, droplets (20  $\mu$ l) of conidium suspension ( $1 \times 10^6$ – $1.5 \times 10^6$  conidia/ml in distilled water) were inoculated on an inductive surface of projection membrane (Gaoke, China) for 23.5–24.5 h at 25 °C. And then appressoria were harvested and from which total RNA was isolated with Trizol (Molecular Research Center, USA) following the manufacturer's procedure.

### Appressorium cDNA synthesis and library construction

The ss cDNA, ds cDNA and library construction were carried out according to Smart cDNA Library Construction Kit User Manual (CLI, 2001). Three  $\mu$ l (~5.23  $\mu$ g) total RNA was used to reverse transcribe to single-strand cDNA with SMART IV<sup>TM</sup> Oligonucleotide, CDS III/3' PCR Primer and PowerScript<sup>TM</sup> Reverse Transcriptase at 42 °C for 1 h. Two  $\mu$ l of first-strand cDNA was used to carry out ds cDNA synthesis by 23 cycles of PCR (95 °C for 15 s, 68 °C for 6 min) with 5' PCR primer, CDS III/3' PCR primer and Advantage<sup>TM</sup> 2 polymerase mix. Half of the ds cDNA were digested with proteinase K and purified with phenol:chloroform: isoamylalcohol. Then all purified ds cDNA was digested with Sfi I enzyme. The reaction product was fractionated by CHROMA SPIN-400 column and the first three fractions containing ds cDNA were collected. The cDNA of all three fractions containing ds cDNA was concentrated by ethanol and resuspended in 7  $\mu$ l deionized H<sub>2</sub>O. At last, these Sfi I-digested cDNAs were ligated to Sfi I-digested, dephosphorylated  $\lambda$ TriplEx2 vector at 16 °C overnight. The ligated cDNA/ $\lambda$ TriplEx2 vector was packed using MaxPlax<sup>TM</sup> Lambda Packaging Extracts (Epicentre, USA).

### Analysis of the appressorium cDNA library

The unamplified cDNA library was titered and the percentage of recombinant clones was determined by blue/white screening according to the instructions in manual of SMART<sup>TM</sup> cDNA library construction kit (Clontech, USA).

Twelve randomly selected plaques were used for the conversion of  $\lambda$ TriplEx2 clone to pTriplEx2 plasmid according to the instructions of manual of SMART<sup>TM</sup> cDNA library construction kit. The plasmids were purified by CTAB-method DNA Mini Preparation and digested by Sfi I enzyme. The inserts

were analyzed by separation on a 1% agarose/EtBr gel. And the expressed sequence tags (EST) of nine randomly selected clones were sequenced with T7 primer on an ABI 377 DNA sequencer. The sequence data were analyzed using VecScreen program (NCBI) for vector masking, and then the adaptor sequences were removed from these cDNA sequences. These EST sequences were processed using software BioEdit (Hall, 1999) for contig assembly. Process sequences were subjected to similarity searches against GenBank database using Blast 2.2.8 (Altschul et al., 1997) and against Phytopathogenic Fungi and Oomycete EST database (Version 1.4) in COGEME (<http://cogeme.ex.ac.uk/index.html>) and *Magnaporthe grisea* database (<http://www.broad.mit.edu/annotation/fungi/magnaporthe/>).

### Appressorium cDNA library amplification and titer of the amplified library

The manual of SMART<sup>TM</sup> cDNA library construction kit suggested amplification of the unamplified cDNA library and then titering of the amplified library.

## RESULTS

### Appressorium induction of *M. grisea*

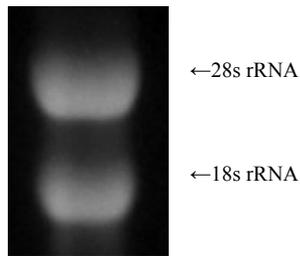
More than 96% conidia inoculated on the hydrophobic surface of a projection membrane for 23.5–24.5 h at 25 °C germinated and formed appressoria (Fig.1).



Fig.1 Appressoria of *M. grisea* GUY11 on a projection membrane inoculated for 23.5–24.5 h at 25 °C

### Total RNA isolation of appressorium

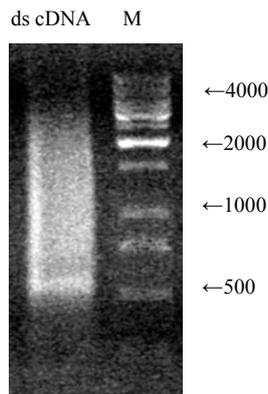
The ratio of OD<sub>260</sub>/OD<sub>280</sub> of total RNA isolated from appressoria incubated for 23.5–24.5 h was 1.63. From the gel, total RNA appeared as two bright bands (28s rRNA and 18s rRNA) and showed intact (Fig.2).



**Fig.2** Appressorium total RNA (5  $\mu$ l) on a 1.2% denaturing formaldehyde/agarose gel

### cDNA synthesis and library construction quality analysis

The sizes of the ds cDNA produced from the 24 h appressorium mRNA ranged from 200 bp to 6 kb, mainly between 500 bp and 5 kb on a 1.1% agarose gel (Fig.3). After fractionation, the first three fraction containing cDNA (8, 9 and 10 fractions) were collected and used for ligation to vector.

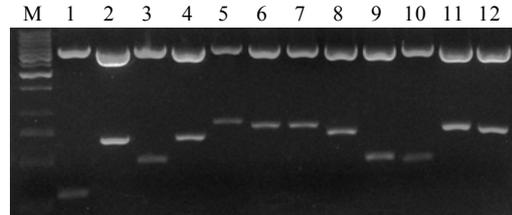


**Fig.3** Ten  $\mu$ l ds cDNA product synthesized from appressorium RNA electrophoresed on a 1.1% agarose/EtBr gel Lane M, 1-kb DNA size marker (Fermentas, Lithuania) (0.2  $\mu$ g loaded)

The cDNA was ligated unidirectionally into  $\lambda$ TriplEx2 vector by three ligation reactions. From the three ligations combined, a primary library of  $2.37 \times 10^6$  independent clones was obtained. The result of blue/white screening of the primary cDNA library in *E. coli* XL1-Blue showed that the ratio of white (recombinant) to blue (nonrecombinant) in two LB/MgSO<sub>4</sub> plates containing IPTG and X-gal were 684/0 and 931/0 respectively. So, the recombination efficiency of cDNA ligation reached 100%. After amplification, the amplified cDNA library had a titer of  $2.86 \times 10^9$  pfu/ml.

Twelve randomly selected lambda plaques of the primary library were converted to plasmids by in vivo

conversion and were analyzed for insert size. The plasmids were purified with CTAB method and digested by Sfi I. All 12 plasmids showed that the insert sizes ranged from 300 bp to 900 bp, average of 660 bp (Fig.4).



**Fig.4** Sfi I-digested plasmids of 12 clones on a 1% agarose/EtBr gel

Lane M, 1-kb DNA marker size (Fermentas, Lithuania). 1~12: clones 1~12

Nine randomly selected cDNA clones were partially sequenced using T7 primers. Analysis of the 9 cDNA sequences using BioEdit software (Hall, 1999) generated 8 nonredundant EST sequences. All 8 unique EST sequences were subjected to similarity searches against GenBank (Blastn 2.2.8 and Blastx 2.2.8) (Altschul *et al.*, 1997), COGEME Db and *Magnaporthe grisea* Database. Six of them had homologous sequences of *M. grisea* in dbEST and the other 2 clones' EST sequences were found to be inhomologous sequences (Table 1) until submission. Clone C02 was homologous to putative protein of cDNA of *Neurospora crassa* (GenBank Accn: AW723279, Expect value=3e-53). As for clone C04, no homologous putative protein sequence or EST sequence hits with *E*-value less than  $e^{-3}$  were found in NCBI blast. These sequences have been deposited in the dbEST of GenBank database (GenBank Accn: CK725218~CK725225).

**Table 1** Homologous sequences from *M. grisea* of 8 EST sequences by Blastn in GenBank

Clone noes	Putative product/function
C01	Glutathione peroxidase (BU641649, $e^{-140}$ )*
C02	Not found
C04	Not found
C05	Cyclophilin (peptidyl-prolyl cis-trans isomerase) (BI808723, 0.0)
C06	ATP synthase subunit 9, lipid-binding protein, mitochondrial (AA415145, 0.0)
C09	Unknown (CD031047, 0.0)
C11	Unknown (CD036090, $e^{-119}$ )
C12	Unknown (BM872180, 0.0)

\* GenBank accession numbers and *E*-values for homologues are in parentheses

## DISCUSSION

Although there are many *M. grisea* cDNA libraries in the world, mainly in Ebbole's lab at Texas A&M University and Ralph Dean's lab at North Carolina State Biotechnology Center (Ebbole *et al.*, 2002; Xu and Xue, 2002), and there are five reported appressorium cDNA libraries built from germinated conidia on appressorium-inductive surface ([http://www.tigr.org/tdb/tgi/mggi/searching/xpress\\_search.html](http://www.tigr.org/tdb/tgi/mggi/searching/xpress_search.html)), no cDNA library was built strictly from mature appressorium. The appressorium cDNA lambda expression library reported here was constructed from mature appressoria inoculated on an inductive surface for 23.5~24.5 h at 25 °C, and had  $2.37 \times 10^6$  pfu independent clones with 100% recombinants enough to screen genes expressed in appressorium. In addition, all cDNA sequences inserted into MCS of  $\lambda$ TriplEx2, the vector of this library, can be expressed for screening genes with protein antigens. And the vector  $\lambda$ TriplEx2 is easily converted from phage to a plasmid vector via Cre-lox-mediated subcloning.

Two of 8 non-redundant EST sequences have no homologues with *E*-value less than  $e^{-3}$  in *M. grisea* ESTs available in GenBank until submission. This cDNA library probably contains many genes missed by previous *M. grisea* cDNA libraries. Clone C05 encoded a reported peptidyl prolyl cis-trans isomerase (cyclophilin). Cyclophilin gene *CYP1* is a virulence factor in *M. grisea* (Viaud *et al.*, 2002). *CYP1* mutants show reduced virulence and are impaired in associated functions, such as penetration peg formation and inducement of appressorium turgor. From this cDNA library made from mature appressoria, a 3.0 kb long cDNA sequence of *MGT1* gene has been successfully cloned (<sup>1</sup>Wang, unpublished). The appressorium cDNA library ESTs can be used for identification of genes expressed in appressorium. For example, promoters of 12 genes were isolated from the rice blast fungus based on the sequences of randomly ESTs (appressorium formation stage cDNA library of *Magnaporthe* available from GenBank) and then these promoters can be used to identify the genes expressed during infection stages (Banno *et al.*, 2003).

So, this 24 h appressorium cDNA library is a representative cDNA library that reveals the gene expression state in appressorium and is suitable for gene expression and function analysis of *M. grisea* appressorium.

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