



## Review

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# Prefrontal cortical circuits in social behaviors: an overview

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**Abstract:** Social behaviors are fundamental and intricate functions in both humans and animals, governed by the interplay of social cognition and emotions. A noteworthy feature of several neuropsychiatric disorders, including autism spectrum disorder (ASD) and schizophrenia (SCZ), is a pronounced deficit in social functioning. Despite a burgeoning body of research on social behaviors, the precise neural circuit mechanisms underpinning these phenomena remain to be elucidated. In this paper, we review the pivotal role of the prefrontal cortex (PFC) in modulating social behaviors, as well as its functional alteration in social disorders in ASD or SCZ. We posit that PFC dysfunction may represent a critical hub in the pathogenesis of psychiatric disorders characterized by shared social deficits. Furthermore, we delve into the intricate connectivity of the medial PFC (mPFC) with other cortical areas and subcortical brain regions in rodents, which exerts a profound influence on social behaviors. Notably, a substantial body of evidence underscores the role of *N*-methyl-D-aspartate receptors (NMDARs) and the proper functioning of parvalbumin-positive interneurons within the mPFC for social regulation. Our overarching goal is to furnish a comprehensive understanding of these intricate circuits and thereby contribute to the enhancement of both research endeavors and clinical practices concerning social behavior deficits.

**Key words:** Prefrontal cortex (PFC); Social behavior; Autism spectrum disorder (ASD); Schizophrenia (SCZ); Parvalbumin-positive interneuron; *N*-Methyl-D-aspartate receptor (NMDAR)

## 1 Introduction

Generally, social behaviors can be defined as any form of communication and interaction between two conspecifics. These behaviors are indispensable and complex in many species and essential for survival and reproduction (Ebstein et al., 2010; McGraw and Young, 2010). However, when exhibited at inappropriate time, places, or intensity, social behaviors can have detrimental effects on both individuals and the social group as a

whole. Impairments in social functioning are prominent features of several neuropsychiatric disorders, including autism spectrum disorder (ASD) and schizophrenia (SCZ) (Chen and Hong, 2018).

The prefrontal cortex (PFC) and its extensive bidirectional connections, forming a top-down modulatory system for social behaviors, are components of the intricate and vast neural networks related to social behaviors (Anastasiades and Carter, 2021; Klune et al., 2021). Throughout the evolution of mammals, the prefrontal region has expanded in size relative to the rest of the cortex, reaching its largest volume in the human brain. Present-day neuroscientists have increasingly turned to research involving mice to gain insights into the underlying mechanisms taking place in the PFC for social behaviors. Studies in both humans and

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animals have underscored the central role of the frontal brain regions in cognition (Carlén, 2017).

Despite the rapid advancements in molecular, cellular, and genetic methodologies, alongside the implementation of cutting-edge imaging technologies, the precise neural circuit mechanisms responsible for social behaviors remain elusive. In this paper, we aim to provide a concise review of the role of PFC in mediating a wide range of social behaviors in ASD/SCZ patients and animal models.

## 2 Prefrontal cortex and social regulation in humans

### 2.1 Structure and function of the PFC

The PFC plays an indispensable role in higher brain functions, including cognition, motivation, reward, and emotion. It also governs goal-directed behaviors, social behaviors, and moral judgement (Forbes and Grafman, 2010; Buschman and Miller, 2014; Hanganu-Opatz et al., 2023). In humans, the PFC can be divided into distinct regions, including dorsolateral PFC (dlPFC), ventrolateral PFC (vlPFC), dorsomedial PFC (dmPFC), ventromedial PFC (vmPFC), and orbitofrontal cortex (OFC) (Forbes and Grafman, 2010). While the medial PFC (mPFC) regions are specifically associated with social behaviors, the lateral regions, including the dlPFC and vlPFC, may become active during social tasks while they are generally regarded as “domain general” (Amodio and Frith, 2006; Mitchell et al., 2006). Other brain regions commonly recruited by social behaviors in humans include the anterior cingulate cortex (ACC), nucleus accumbens (NAc), amygdala, hippocampus (HPC), superior temporal sulcus, and temporal parietal junction (Gangopadhyay et al., 2021; Chen et al., 2023). Regions of the PFC exhibit dense interconnections. Significant glutamatergic projections emanate from the PFC to the ACC, thalamus, ventral tegmental area (VTA), HPC, and NAc. Furthermore, glutamatergic neurons arise from the HPC and innervate the hypothalamus (HT), VTA, NAc, and PFC, and from the amygdala to HT, ACC, and NAc (Sarawagi et al., 2021). From birth to early adulthood, PFC cells and circuits undergo physiological changes, altering their strength of connectivity with distant brain regions. The extended maturation of the PFC likely enables the emergence of complex behaviors

but may render them more susceptible to disruptions (Kolb et al., 2012; Klune et al., 2021).

Studies in humans have highlighted the central role of the frontal brain regions in cognition. Higher-order cognitive abilities encompass attention, salience detection, working memory, strategy shifting, and inhibitory control, all of which facilitate adaptation to varying conditions (Diamond, 2013; Buschman and Miller, 2014). These abilities necessitate the coordination of cellular ensembles that balance the stability and flexibility of neural representations. During baseline activity, prefrontal neurons lack the functional coordination of activity; however, when task-related demands arise, the temporal coordination of activity dynamically binds prefrontal neurons into functional units through oscillations and polychrony (Hanganu-Opatz et al., 2023). This synchronization influences communication between neuronal groups and ensures that presynaptic activation patterns reach postsynaptic neurons in a temporally coordinated manner. Oscillatory phase locking is primarily driven by bottom-up-directed gamma band activity (30–90 Hz) (Fries, 2015). In each oscillatory cycle, representing a transition between an averaged depolarized state and a hyperpolarized state, neurons have a temporal window to synchronize their firing, establishing connections (Buzsáki and Draguhn, 2004). Oscillations are hypothesized to create a temporal scaffold for intra- and inter-brain area communication (Buzsáki and Llinás, 2017). The gamma rhythm primarily arises from the synchronized fast inhibition of excitatory neurons by parvalbumin (PV)-expressing interneurons, demonstrating its role as a resonant property in local networks (Cardin et al., 2009; Sohal et al., 2009). It has been suggested that local gamma oscillations mirror the transient activation of PFC neuronal assemblies (Fujisawa et al., 2008). The gamma rhythm usually occurs along with the theta rhythm (4–12 Hz). The synchronization of theta rhythm between the PFC and HPC can influence various cognitive states, including memory integration and sequential working memory (Backus et al., 2016; Su et al., 2024). To this end, research has illuminated the vital role of oscillatory synchrony, particularly gamma and theta rhythms, in facilitating communication within the PFC and among various brain areas, which underscores their significance in orchestrating complex behaviors, including social interactions.

## 2.2 Cognitive and emotional processing in social regulation

Social behavior deficits represent a fundamental dimension of many psychiatric disorders, such as the neurodevelopmental disorders, ASD and SCZ. In social decision-making contexts, individuals frequently need to recognize other individuals, infer their intentions and emotions, and weigh the values of both social and non-social outcomes before choosing an action. To a significant extent, these aspects of social information processing rely on the PFC, specifically the mPFC (Kietzman and Gourley, 2023). Consequently, social cognition and emotion play critical roles in social regulation.

Social cognition encompasses a wide array of mental operations used for identifying and interpreting social signals, as well as the utilization of these signals to flexibly guide appropriate social behaviors in changing environments (Millan and Bales, 2013). Social cognition has three major facets: social motivation, self and other knowledge, and group dynamics. Research using both humans and translational animal models has demonstrated that these aspects of social cognition relate to psychiatric disorders. Social motivation can be described as biasing the individual to preferentially orient to the social world (social orienting), to seek and take pleasure in social interactions (social reward), and to strive to foster and maintain social bonds (social maintaining). The social motivation theory of autism suggests that a lack of early social interest may contribute to subsequent social cognitive deficits that manifest themselves later in development (Chevallier et al., 2012). Thus, social motivation can be seen as a developmental and evolutionary foundation for other social behaviors. Self and other knowledge forms the core of social cognition and encompasses concepts explored in basic processes like facial recognition, action perception, and empathy, as well as more sophisticated social behaviors such as cooperation and intergroup interaction (Catmur et al., 2016). Living in groups is common in mammalian societies, and dominance hierarchy is a fundamental organizing mechanism based on repeated social competitions. The result of social competition is influenced by personality traits under the regulation of high cortical functions (Zhou et al., 2018). Among the high cortical regions, the mPFC has been specifically associated with social dominance (Wang et al., 2011; Zhang

et al., 2022; Fan et al., 2023). Hence, social cognition encompasses a wide range of mental processes crucial for our understanding of social interactions. These facets of social cognition not only have been linked to psychiatric disorders in humans and animal models but also offer insights into the fundamental building blocks of social behaviors.

Emotion regulation involves processes that affect both the experience and expression of emotions, which plays a crucial role in facilitating successful social interactions and overall functioning (McRae and Gross, 2020). Effective emotion regulation is correlated with an enhanced capacity to engage in socially appropriate emotions and behaviors, contributing to adaptive social interactions and social competence (Eisenberg et al., 1995, 2000). Conversely, disruptions in effective emotion regulation strategies may arise from negative social experiences, leading to difficulties in social behaviors and negative social encounters (Eisenberg et al., 1999; Geckeler et al., 2022). These intricate processes of emotion regulation play pivotal roles in shaping the quality of social interactions and overall social functioning.

Taken together, the PFC, with its intricate network of regions and dynamic neural activity patterns, serves as the cornerstone of higher brain functions, influencing cognition, emotions, social behaviors, and more. At the same time, addressing social behavior deficits in psychiatric disorders like ASD and SCZ necessitates a comprehensive exploration of the interconnected dimensions of social cognition and emotion regulation. These conditions often involve complex processes of social information processing that heavily rely on the PFC, particularly the mPFC.

## 3 Prefrontal dysfunction in autism and schizophrenia

### 3.1 Brain imaging as evidence for impaired PFC networks in ASD

Numerous researchers have underscored the pivotal role of the PFC in the manifestation of various ASD symptoms. For instance, functional magnetic resonance imaging (fMRI) studies have revealed that dysfunctional networks within the frontal lobes may affect social cognition, repetitive behavior, executive

function, and verbal communication—functions primarily controlled by the frontal lobe (Baron-Cohen et al., 1999; Carper and Courchesne, 2000, 2005; Ohnishi et al., 2000; Luna et al., 2002; Herbert et al., 2003; Chandana et al., 2005; Rafiee et al., 2022). Moreover, individuals with ASD have been observed to exhibit diminished serotonergic synthesis in the frontal lobes, alongside concomitant abnormalities in global brain serotonin production (Chugani et al., 1999; Chandana et al., 2005). Serotonin, similar to the brain-derived neurotrophic factor (BDNF), plays a critical role in modulating axonal arborization. Dysregulation of serotonin synthesis has been linked to abnormalities in frontal lobe connectivity (Kumar et al., 2010).

Disorders of neural connectivity, which explain a substantial portion of ASD variance, enable the integration of cognitive theories (e.g., central coherence) with a comprehensive understanding of ASD behaviors at the neuroanatomical, neurophysiological, and neuropsychological levels (Hughes, 2007, 2008). Impaired integration in ASD may arise from functional dysconnectivity among brain systems. Existing data suggest both cortico-subcortical and inter-/intra-hemispheric hyperconnectivity or hypoconnectivity (Vissers et al., 2012; Uddin et al., 2013; Kleinhans et al., 2016; Martínez et al., 2020). The function of frontal streams, as presented in the following section, has been validated through postmortem studies conducted on individuals with ASD. An important finding was the observation of brain overgrowth postpartum early in life, particularly of the PFC, which correlated with significantly larger head circumferences in children under three years of age (Courchesne et al., 2003; Sacco et al., 2015). Subcortical brain regions involved in sensory processing including the cerebellum and thalamus, both of which project to the PFC, exhibit anatomical differences in comparison to neurotypical individuals (Hazlett et al., 2005; Courchesne et al., 2011a, 2011b). The underconnectivity observed between frontal-thalamic areas has been associated with the severity of symptoms in individuals with ASD (Martínez-Sanchis, 2014; Rane et al., 2015). Moreover, there are strong connections between the PFC and the cerebellum, and abnormalities in both regions have been linked to symptom severity (Carper and Courchesne, 2000; Kumar et al., 2010).

### 3.2 Brain imaging as evidence for impaired PFC networks in SCZ

Numerous reports have underscored the significant similarities between the cognitive deficits observed in patients with frontal lesions and those encountered in SCZ (Müller et al., 2002). Furthermore, functional imaging studies have shed light on altered PFC activation during cognitive testing in SCZ patients, supporting the hypothesis that disruptions in PFC activity cause deficits in working memory and other cognitive functions (Weinberger and Berman, 1996; Karlsgodt et al., 2009). In fact, SCZ patients exhibit reduced blood flow in the PFC during task performance, along with diminished functional connectivity between the PFC and other brain regions while engaged in cognitive tasks (Goldman-Rakic and Selemon, 1997; Fornito et al., 2009).

A crucial source of excitatory input to the PFC originates from the thalamus, notably the mediodorsal nucleus. A reduction in correlated activity between the thalamus and the PFC during cognitive assessments could be a potential factor contributing to cognitive deficits (Mitelman et al., 2005; Woodward et al., 2012; Giraldo-Chica et al., 2018). Similarly, assessments of resting-state functional connectivity have indicated reduced thalamo-prefrontal connectivity in various stages of SCZ, including in adolescents with early-onset SCZ, young individuals at clinical high risk, and adults in the initial phases of the disorder (Anticevic et al., 2015; Cho et al., 2016; Woodward and Heckers, 2016; Huang et al., 2021; Zhang et al., 2021). Notably, the reduction in thalamo-prefrontal connectivity among high-risk individuals becomes the most pronounced in those who are subsequently diagnosed with the disorder (Anticevic et al., 2015). These findings intriguingly suggest that thalamo-PFC dysconnectivity during adolescence might contribute to the developmental etiology of SCZ (Anticevic et al., 2015; Woodward and Heckers, 2016).

SCZ is associated with both atypical PFC development during embryonic stages and impaired PFC maturation in later adolescence, influenced by environmental stressors. Due to this unique aspect, SCZ can be characterized by two critical susceptibility periods (Selemon and Zecevic, 2015). The disruption of cortical synaptic pruning has been proposed as a key factor in the etiology of SCZ, given that its symptoms typically manifest themselves in late adolescence,



which coincides with the developmental period when PFC connectivity matures (Feinberg, 1990; Lewis, 1997). Gray matter loss, reflecting cortical maturation, tends to occur later in higher-order association cortices such as the prefrontal and temporal cortices compared with lower-order somatosensory and visual cortices, indicating a delayed maturation of these cortical regions (Gogtay et al., 2004). Cortical gray matter loss is more pronounced in individuals at clinical high risk for SCZ who later develop the disorder, in contrast to others (Cannon et al., 2015; Chung et al., 2017). These observations suggest that alterations in prefrontal maturation may play a role in the etiology of the disorder.

Findings from postmortem studies have consistently revealed changes in elements of GABAergic neurotransmission within the PFC of individuals with SCZ. The effectiveness of GABAergic neurotransmission correlates with the availability of  $\gamma$ -aminobutyric acid (GABA) in the synapse. In SCZ, both messenger RNA (mRNA) and protein levels of 67 kDa glutamic acid decarboxylase (GAD67), the primary enzyme responsible for cortical GABA synthesis, are diminished in the PFC (Vawter et al., 2002; Woo et al., 2008; Curley et al., 2011; Kimoto et al., 2014). This aligns with the notion that the availability of presynaptic GABA is diminished in this disorder. Additionally, the numbers of PV<sup>+</sup> (Beasley and Reynolds, 1997; Beasley et al., 2002; Hashimoto et al., 2003), cholecystokinin-positive (CCK<sup>+</sup>) (Hashimoto et al., 2008; Fung et al., 2010), and somatostatin-positive (SST<sup>+</sup>) (Hashimoto et al., 2008; Morris et al., 2008; Fung et al., 2010) interneurons are decreased in the PFC of individuals with SCZ. Interestingly, GABA concentration has been found to predict the gamma oscillatory frequency (Edden et al., 2009). There is substantial evidence supporting *N*-methyl-D-aspartate receptor (NMDAR) hypofunction in SCZ, as the administration of NMDAR antagonists, such as MK-801 or phencyclidine, could induce SCZ-like symptoms in human subjects. Consequently, it is postulated that the NMDAR hypofunction in SCZ primarily affects cortical GABAergic interneurons (Nakazawa et al., 2017).

In brief, mounting evidence regarding PFC impairment in both ASD and SCZ patients highlights the critical role of this brain region in the manifestation of symptoms in these disorders. Brain imaging studies have unveiled disrupted PFC and PFC network

functionality, enhancing our understanding of the cognitive deficits and behavioral challenges seen in these conditions. Additionally, insights from studies examining neural connectivity and neurotransmission further emphasize the significance of the PFC in the pathophysiology of ASD and SCZ. Understanding these neural underpinnings is crucial for developing targeted interventions and treatments to improve the lives of individuals affected by these disorders.

## 4 Prefrontal mechanisms of social deficits in rodents

### 4.1 Measuring social behaviors in rodents

A multitude of studies have demonstrated that rodents exhibit behaviors closely aligned with ethological aspects of social processing demands. These behaviors encompass sociability, social memory/recognition, and dominance, offering conceptual parallelism with human social categories (Moy et al., 2004; Bariselli et al., 2018; Zhou et al., 2018). Given their inherent social nature, mice serve as invaluable research models, facilitating the exploration of aberrant social behaviors associated with ASD and SCZ.

In order to assess sociability in mice, behavioral paradigms like social preference tests have often been employed. These tests assess the amount of time spent in proximity to an unfamiliar mouse compared with an empty side within a three-chamber apparatus. Typically, the novel social stimulus is confined to a compartment that allows sniffing and interaction but prevents physical contact (Moy et al., 2004). Alternatively, sociability can be evaluated by measuring the time spent engaged in unconstrained interactions.

In order to quantify social recognition in mice, researchers can measure the time spent with novel versus familiar conspecifics in a three-chamber apparatus (Moy et al., 2004). Typically, wild-type mice exhibit a preference for social novelty by spending more time with the novel conspecifics. Other paradigms for measuring social recognition take advantage of mice's natural tendency to habituate to familiar conspecifics while showing greater interest in exploring a novel mouse. A reduction in sniffing time during repeated trials indicates recognition of the familiar mouse. After repeated presentations, a novel mouse is introduced, and an increased investigation of the novel

mouse reflects a preference for social novelty (Bariselli et al., 2018).

Dominance hierarchy serves as a fundamental organizing mechanism in most animal societies. Once established, the hierarchical rank remains relatively stable and reduces intense conflicts among group members. To assess the dominance tendencies of mice, the tube test was developed, in which one mouse, termed as “loser,” is compelled to back out of a narrow tube by another mouse, referred to as “winner” (Wang et al., 2011; Zhang et al., 2022). A simpler method is to observe the behaviors of animals within their home cages or to monitor aggressive interactions, which typically occur when a group is placed into a new cage (Shemesh et al., 2013).

Play behavior can be observed in various animals, including rodents. In rats, most play behaviors are social, encompassing play fighting, where individuals compete for an advantage (Palagi et al., 2016; Pellis et al., 2023). Play fighting activity peaks during the juvenile period, with juveniles exhibiting a strong motivation to engage peers in such interactions (Varlinskaya et al., 1999). Consequently, social play interactions serve as a primary source of social experience during this developmental stage. It is important to note, however, that play fighting does not specifically train particular motor actions; instead, it enhances a skill set applicable in various social and non-social contexts. Depriving juvenile rats of typical peer-to-peer play experiences results in adults displaying socio-cognitive deficiencies, which correlate with physiological and anatomical alterations in neurons, notably in the PFC, especially the mPFC (Pellis et al., 2023).

Over the last decade, there has been a surge of investigations focused on how mPFC circuits and pathways modulate social behaviors. While the hypothalamus and other subcortical circuits play a central role in generating social behaviors, the mPFC is uniquely equipped to integrate pertinent information such as social rank, memories, and contextual factors to modulate these behaviors. In the subsequent sections, we review the available evidence indicating that activity within the rodent mPFC can lead to alterations in social behaviors.

## 4.2 Involvement of the mPFC in social deficits in rodents

The mPFC of rodents consists of the ACC, the prelimbic cortex (PL), and the infralimbic cortex (IL),

each of them with unique connectivity and functional properties. Multiple lines of evidence indicate the significance of neural activity originating from the mPFC in shaping social behaviors in rodents. Although the medial aspect of the secondary motor cortex (M2) is sometimes considered to be a part of the rodent mPFC, we do not include it in this review (Barthas and Kwan, 2017).

### 4.2.1 Altered synaptic transmission in the mPFC

Studies involving rodents have shown that dysfunction in the PFC reduces social interest (Murray et al., 2015), impairs social memory (Rudebeck et al., 2007), hinders the processing of social hierarchy information, and lowers social rank (Wang et al., 2014). Research has also delved into synaptic transmission in mouse models with genetic modification derived from risk genes for ASD and SCZ, and revealed general synaptic dysfunctions in the cortex and in some cases, specifically in the PFC. These studies have involved the abnormal expression of various molecules, including methyl-CpG-binding protein 2 (MeCP2) (Sceniak et al., 2016),  $\beta$ 2-subunit nicotinic receptor (Avale et al., 2011), insulin receptor substrate protein, 53 kDa (IRSp53) (Chung et al., 2015), SH3 and multiple ankyrin repeat domains protein 3 (Shank3) (Duffney et al., 2015; Lee et al., 2015), and neuroligin 3 (NL3) (Cao et al., 2018), which have all been linked to altered synaptic transmission in the mPFC and abnormal social behavior.

Excitation/inhibition imbalance and synaptic dysfunction have been identified as common pathophysiological features in various mouse models of ASD and SCZ. For instance, the conditional knockout of neuroligin 2 (NL2) in the adult mPFC induced significant reductions in synaptic inhibition and caused parallel impairments in anxiety, fear memory, and social interaction behaviors (Liang et al., 2015). Also, NMDAR hypofunction and dysfunction of PV<sup>+</sup> interneurons have been observed in NL3-deficient mice, and the micro-infusion of NMDAR co-agonist D-cycloserine (DCS) into the mPFC rescued the social deficits in these mice (Cao et al., 2022). Similar findings have been reported in other mouse models. For example, a 16p11.2 deletion mouse model exhibited NMDAR hypofunction in PFC pyramidal neurons and cognitive and social impairments, which could be rescued by the chemogenetic activation of PFC pyramidal neurons

(Wang et al., 2018). Moreover, treatment with NMDAR antagonists such as dizocilpine (MK-801) or phencyclidine could induce SCZ-like symptoms in rodents. These symptoms encompassed hyperlocomotion, indicative of positive symptoms, as well as deficits in social behavior observed during a social interaction test and increased immobility in a forced swimming test, serving as indices of negative symptoms (Kehrer et al., 2008; Nakazawa et al., 2017; Kruse and Bustillo, 2022). This strongly supports the NMDAR hypofunction hypothesis of SCZ. Overall, the above findings suggest the importance of normal synaptic function in the mPFC, particularly the role of NMDAR, and provide valuable insights into potential therapeutic approaches targeting specific molecular and synaptic mechanisms, such as NMDARs, for the development of more effective treatments for ASD and SCZ.

Excitation/inhibition imbalance within the mouse mPFC has been linked to profound social deficits (Yizhar et al., 2011). Studies of mouse models lacking contactin-associated protein-like 2 (*CNTNAP2*) gene, implicated in autism, have demonstrated that modulating excitation/inhibition in the mPFC through neuron type-selective, real-time, reversible optogenetic manipulations can acutely rescue deficits in social behavior and hyperactivity (Selimbeyoglu et al., 2017). Moreover, a 16p11.2 duplication mouse model exhibited deficient GABAergic synaptic transmission and elevated excitability in PFC pyramidal neurons, as well as social and cognitive deficits. These deficits could be restored by the expression of neuronal PAS domain-containing protein 4 (*Npas4*), a key regulator of GABA synapses, in PFC (Rein et al., 2021). In line with this finding, it has been commonly observed that cortical inhibition and/or PV<sup>+</sup> interneuron number decreased in mouse models with social impairments (Han et al., 2012; Filice et al., 2016; Cao et al., 2018, 2022). These results indicate that maintaining normal synaptic transmission and proper inhibition within the mPFC neuronal network is essential for normal social behaviors.

#### 4.2.2 Neuronal oscillations in the mPFC during social interactions

The neural activity of principal neurons in the mPFC is crucial for the social behavior of rodents. Studies have shown that certain mPFC principal neurons increase their activity during social interactions in mice (Lee et al., 2016; Brumback et al., 2018). Calcium

activity recordings in mPFC principal neurons have identified distinct and dynamic ON and OFF neural ensembles that encode social exploration and linked dysfunctions in the activity of these ensembles to abnormal social exploration (Liang et al., 2018). Furthermore, the transient excitation of excitatory neurons in the mPFC through optogenetic manipulation led to reduced sociability (Yizhar et al., 2011) but improved social competition (Zhang et al., 2022).

In addition, neural oscillations in the low gamma frequency range (30–50 Hz) have been found to be disturbed in ASD and SCZ (Gandal et al., 2012; David et al., 2016). Optogenetic studies have revealed that activating PV<sup>+</sup> interneurons is both necessary and adequate for inducing gamma oscillations in the cortex, thereby influencing cortical information processing (Cardin et al., 2009; Sohal et al., 2009). Modulating these oscillations, especially through patterned optogenetic stimulation (40 Hz nested at 8 Hz) of mPFC PV<sup>+</sup> interneurons, has been shown to rescue social deficits in an NL3-deficient mouse model of autism (Cao et al., 2018). Interestingly, the excitation of vasoactive intestinal polypeptide-positive interneurons or the inhibition of PV<sup>+</sup> interneurons induced winning, and vice versa (Zhang et al., 2022). The optogenetic synchronization of either PV<sup>+</sup> interneurons or somatostatin-positive interneurons at a low gamma frequency improved sociability in wild-type mice (Liu et al., 2020). These findings underscore the close correlation between social interaction and elevated gamma rhythms in the prefrontal local field potentials, controlled by PV<sup>+</sup> interneurons. As a result, targeted interventions aimed at restoring neuronal oscillations hold great promise for future therapeutic strategies in neurodevelopmental and neuropsychiatric disorders.

#### 4.2.3 Distinct mPFC circuits involved in social behaviors

The mPFC receives a variety of long-range inputs and sends diverse outputs to many other brain regions, connected via local circuits involving different excitatory and inhibitory neurons and communicating with each other based on specific wiring rules.

Cell type-specific optogenetic and chemogenetic manipulations, behavioral testing, and electrophysiological recordings have been used to investigate the functional coupling of the mPFC with subcortical systems in social behavior. Studies have shown that

projection neurons of the basolateral amygdala (BLA) in the IL are preferentially activated in response to a social cue as compared with BLA-projecting neurons in the PL, and the chemogenetic activation of PL-BLA or the inhibition of IL-BLA circuits impairs social behavior (Huang et al., 2020). Moreover, the activation of BLA input to the mPFC leads to an increase in anxiety-like behavior and a decrease in sociability (Felix-Ortiz et al., 2016). Interestingly, PL-to-NAc stimulation decreases the preference for a social target (Murugan et al., 2017). The ventral HPC (vHPC) has been implicated in social memory, and the inhibition of direct projections from vHPC to mPFC impaired social memory expression (Okuyama et al., 2016; Sun et al., 2020). In addition, the firing rates of VTA dopamine neurons projecting to the mPFC were dramatically decreased in a mouse model of repeated social defeat-induced depression. The activation of VTA projections to the NAc or the inhibition of VTA projections to the mPFC induces susceptibility to social defeat stress (Liu et al., 2018). The mPFC is also involved in processing the affective state of others through non-verbal communication. Utilizing *in vivo* single-cell microendoscopic  $Ca^{2+}$  imaging, researchers have demonstrated that increased synchronous activity of mPFC somatostatin-positive interneurons, which guides the inhibition of pyramidal neurons, has been associated with the discrimination of affective states (Scheggia et al., 2020). Interestingly, prior social experience incentivizes later instrumental choices such as food, and the social incentivization of future choices requires the PL (Kietzman et al., 2022).

Moreover, serotonergic neurons in the dorsal raphe nucleus (DRN) play a role in social behavior modulation through their interaction with the mPFC. The manipulation of synaptic inputs from the mPFC to the DRN influences social avoidance behaviors (Michelsen et al., 2008; Challis et al., 2013; Challis and Berton, 2015). The oxytocinergic system, originating from the hypothalamus and acting through SST<sup>+</sup> interneurons expressing oxytocin receptors (OXTRs) in the mPFC, also has an important function in regulating sociosexual behaviors in female mice but anxiety-related behaviors in male mice (Donaldson and Young, 2008; Nakajima et al., 2014; Li et al., 2016). Furthermore, OXTRs are also expressed on glutamatergic neurons in the mPFC, and the optogenetic stimulation of axons in the BLA arising from

OXTR-expressing neurons in the PFC eliminates the ability to distinguish novel conspecifics from familiar ones (Tan et al., 2019). Thus, this section provides a comprehensive overview of the intricate neural circuits and systems that regulate social behaviors, emphasizing the central role of the mPFC and its interactions with various brain regions and neurotransmitter systems. The mPFC receives inputs from the vHPC, BLA, and VTA, and it sends outputs to the NAc, BLA, VTA, and DRN, all of which are implicated in social memory, sociability, social defeat, or anxiety. The modulation of PFC circuits by monoaminergic systems profoundly affects PFC-mediated social behaviors.

In conclusion, the rodent studies summarized in this section strongly support the notion that PFC neurons and their interconnected neural circuits play a central role in the regulation of social behaviors. Within the microcircuits of the mPFC, NMDARs and PV<sup>+</sup> interneurons have merged as critical components that potentially serve as pivotal elements in the regulation of social behaviors.

## 5 Perspectives

The PFC remains a central focus of inquiry in the field of neuroscience, with the potential to unveil deep insights into the complexities of social behaviors. Deciphering the pivotal role of the PFC in governing social behaviors not only enriches our comprehension of fundamental brain mechanisms but also carries profound implications for addressing social deficits witnessed in psychiatric disorders such as ASD and SCZ.

Future research endeavors are predicted to illuminate the dynamic interplay within the PFC and its impact on our social interactions. There is a necessity for continued exploration into the neural circuitry of the PFC, with the objective of unraveling how this region choreographs a multitude of processes integral to social cognition and emotion regulation. Furthermore, it is essential to approach the neural mechanisms of social activities by considering social networks as a cohesive entity, including the PFC. In rodents, the unique input and output from diverse subregions, layers, and cell types within the PFC govern various facets of social behaviors. For example, neurons in layer 2/3 IL that express the neuropeptide corticotropin-releasing



hormone (CRH) respond to social interactions with familiar over novel mice and release CRH into the rostral lateral septum (rLS) to suppress social interactions with familiar mice through lateral septum (LS) disinhibition (de León Reyes et al., 2023). The mPFC cells that project to the lateral hypothalamus promote dominance behavior during reward competition (Padilla-Coreano et al., 2022). Decreased PV<sup>+</sup> interneuron excitability in the mPFC is a causative factor of social novelty deficits (Cao et al., 2018). Hence, forthcoming research should focus on meticulously categorizing PFC cells according to their precise spatial localization, physical connections, and protein markers. Subsequently, examining the regulatory impact of these smaller cell groupings on social behaviors is imperative. A more comprehensive investigation into the intricate microcirculatory networks within the PFC is also warranted; a deeper understanding of PFC circuits will not only expand our knowledge of social behaviors but also provide new treatment strategies for social disorders.

In the preceding discussion, we emphasized the significance of gamma oscillations in the mPFC concerning social behavior and suggested that targeted interventions aimed at restoring neuronal oscillations might be a promising route for addressing the social disorders of ASD and SCZ. There are three potential approaches for implementing oscillation interventions. The first method involves directly enhancing the weakened oscillations in social behavior through physical interventions. Optogenetics is commonly used in animal studies, but the introduction of exogenous genes into the brain hinders its application in humans. Currently, electrode stimulation based on information decoding (Li et al., 2023) provides a more physiological and precise approach, potentially adaptable for translation in patients if the concern about invasive surgery can be addressed. Additionally, non-invasive neuro-modulations such as transcranial direct current stimulation or transcranial magnetic stimulation can be evaluated in clinical trials, with observations on whether social behavior and gamma oscillation are improved in subjects. The second method comprises pharmacological intervention, utilizing specific drugs that target PV<sup>+</sup> interneurons to regulate their excitability, thereby impacting gamma oscillation. For instance, potential options include drugs targeting voltage-gated sodium channel Nav1.1 (Ogiwara et al., 2007) and voltage-gated

potassium channel Kv3.1 (Kaczmarek and Zhang, 2017), specifically expressed on PV<sup>+</sup> interneurons. Moreover, it is noteworthy that the excitability of PV<sup>+</sup> interneurons can also be modulated by NMDARs (Cao et al., 2022), although this modulation is not exclusive to PV interneurons. The third method entails neuron transplantation. Studies have shown that transplanting Nav1.1-enhanced interneurons can increase gamma oscillation and enhance cognition in a mouse model of Alzheimer's disease (Martinez-Losa et al., 2018). As research progresses, this approach could potentially prove effective in human patients. In summary, the exploration of the above various approaches underscores the potential for targeted interventions in manipulating gamma oscillations, opening up promising avenues for addressing disorders with social dysfunction, such as ASD and SCZ. Continued research in these areas may pave the way for novel therapeutic strategies in the realm of neurobiological interventions for social behavior-related conditions.

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### Author contributions

Jianhong LUO determined the topic of the article, proposed the program, and was responsible for the supervision and manuscript revision. Wei CAO and Huiyi LI were responsible for reference searching and manuscript writing. All authors have read and approved the final manuscript.

### Compliance with ethics guidelines

Jianhong LUO is an Editorial Board Member for *Journal of Zhejiang University-SCIENCE B (Biomedicine & Biotechnology)* and was not involved in the editorial review or the decision to publish this article. Wei CAO, Huiyi LI, and Jianhong LUO declare that they have no conflicts of interest.

This review does not contain any studies with human or animal subjects performed by any of the authors.

### References

- Amodio DM, Frith CD, 2006. Meeting of minds: the medial frontal cortex and social cognition. *Nat Rev Neurosci*, 7(4): 268-277.

- <https://doi.org/10.1038/nrn1884>
- Anastasiades PG, Carter AG, 2021. Circuit organization of the rodent medial prefrontal cortex. *Trends Neurosci*, 44(7): 550-563.  
<https://doi.org/10.1016/j.tins.2021.03.006>
- Anticevic A, Haut K, Murray JD, et al., 2015. Association of thalamic dysconnectivity and conversion to psychosis in youth and young adults at elevated clinical risk. *JAMA Psychiatry*, 72(9):882-891.  
<https://doi.org/10.1001/jamapsychiatry.2015.0566>
- Avale ME, Chabout J, Pons S, et al., 2011. Prefrontal nicotinic receptors control novel social interaction between mice. *FASEB J*, 25(7):2145-2155.  
<https://doi.org/10.1096/fj.10-178558>
- Backus AR, Schoffelen JM, Szebényi S, et al., 2016. Hippocampal-prefrontal theta oscillations support memory integration. *Curr Biol*, 26(4):450-457.  
<https://doi.org/10.1016/j.cub.2015.12.048>
- Bariselli S, Hörnberg H, Prévost-Solić C, et al., 2018. Role of VTA dopamine neurons and neuroligin 3 in sociability traits related to nonfamiliar conspecific interaction. *Nat Commun*, 9:3173.  
<https://doi.org/10.1038/s41467-018-05382-3>
- Baron-Cohen S, Ring HA, Wheelwright S, et al., 1999. Social intelligence in the normal and autistic brain: an fMRI study. *Eur J Neurosci*, 11(6):1891-1898.  
<https://doi.org/10.1046/j.1460-9568.1999.00621.x>
- Barthas F, Kwan AC, 2017. Secondary motor cortex: where 'sensory' meets 'motor' in the rodent frontal cortex. *Trends Neurosci*, 40(3):181-193.  
<https://doi.org/10.1016/j.tins.2016.11.006>
- Beasley CL, Reynolds GP, 1997. Parvalbumin-immunoreactive neurons are reduced in the prefrontal cortex of schizophrenics. *Schizophr Res*, 24(3):349-355.  
[https://doi.org/10.1016/s0920-9964\(96\)00122-3](https://doi.org/10.1016/s0920-9964(96)00122-3)
- Beasley CL, Zhang ZJ, Patten I, et al., 2002. Selective deficits in prefrontal cortical gabaergic neurons in schizophrenia defined by the presence of calcium-binding proteins. *Biol Psychiatry*, 52(7):708-715.  
[https://doi.org/10.1016/s0006-3223\(02\)01360-4](https://doi.org/10.1016/s0006-3223(02)01360-4)
- Brumback AC, Ellwood IT, Kjaerby C, et al., 2018. Identifying specific prefrontal neurons that contribute to autism-associated abnormalities in physiology and social behavior. *Mol Psychiatry*, 23(10):2078-2089.  
<https://doi.org/10.1038/mp.2017.213>
- Buschman TJ, Miller EK, 2014. Goal-direction and top-down control. *Philos Trans Roy Soc B Biol Sci*, 369(1655): 20130471.  
<https://doi.org/10.1098/rstb.2013.0471>
- Buzsáki G, Draguhn A, 2004. Neuronal oscillations in cortical networks. *Science*, 304(5679):1926-1929.  
<https://doi.org/10.1126/science.1099745>
- Buzsáki G, Llinás R, 2017. Space and time in the brain. *Science*, 358(6362):482-485.  
<https://doi.org/10.1126/science.aan8869>
- Cannon TD, Chung Y, He G, et al., 2015. Progressive reduction in cortical thickness as psychosis develops: a multi-site longitudinal neuroimaging study of youth at elevated clinical risk. *Biol Psychiatry*, 77(2):147-157.  
<https://doi.org/10.1016/j.biopsych.2014.05.023>
- Cao W, Lin S, Xia QQ, et al., 2018. Gamma oscillation dysfunction in mPFC leads to social deficits in neuroligin 3 R451C knockin mice. *Neuron*, 97(6):1253-1260.e7.  
<https://doi.org/10.1016/j.neuron.2018.02.001>
- Cao W, Li JH, Lin S, et al., 2022. NMDA receptor hypofunction underlies deficits in parvalbumin interneurons and social behavior in neuroligin 3 R451C knockin mice. *Cell Rep*, 41(10):111771.  
<https://doi.org/10.1016/j.celrep.2022.111771>
- Cardin JA, Carlén M, Meletis K, et al., 2009. Driving fast-spiking cells induces gamma rhythm and controls sensory responses. *Nature*, 459(7247):663-667.  
<https://doi.org/10.1038/nature08002>
- Carlén M, 2017. What constitutes the prefrontal cortex? *Science*, 358(6362):478-482.  
<https://doi.org/10.1126/science.aan8868>
- Carper RA, Courchesne E, 2000. Inverse correlation between frontal lobe and cerebellum sizes in children with autism. *Brain*, 123(Pt 4):836-844.  
<https://doi.org/10.1093/brain/123.4.836>
- Carper RA, Courchesne E, 2005. Localized enlargement of the frontal cortex in early autism. *Biol Psychiatry*, 57(2): 126-133.  
<https://doi.org/10.1016/j.biopsych.2004.11.005>
- Catmur C, Cross ES, Over H, 2016. Understanding self and others: from origins to disorders. *Philos Trans Roy Soc B Biol Sci*, 371(1686):20150066.  
<https://doi.org/10.1098/rstb.2015.0066>
- Challis C, Berton O, 2015. Top-down control of serotonin systems by the prefrontal cortex: a path toward restored socioemotional function in depression. *ACS Chem Neurosci*, 6(7):1040-1054.  
<https://doi.org/10.1021/acchemneuro.5b00007>
- Challis C, Boulden J, Veerakumar A, et al., 2013. Raphe GABAergic neurons mediate the acquisition of avoidance after social defeat. *J Neurosci*, 33(35):13978-13988.  
<https://doi.org/10.1523/JNEUROSCI.2383-13.2013>
- Chandana SR, Behen ME, Juhász C, et al., 2005. Significance of abnormalities in developmental trajectory and asymmetry of cortical serotonin synthesis in autism. *Int J Dev Neurosci*, 23(2-3):171-182.  
<https://doi.org/10.1016/j.ijdevneu.2004.08.002>
- Chen P, Hong WZ, 2018. Neural circuit mechanisms of social behavior. *Neuron*, 98(1):16-30.  
<https://doi.org/10.1016/j.neuron.2018.02.026>
- Chen XL, Liu JX, Luo YJ, et al., 2023. Brain systems underlying fundamental motivations of human social conformity. *Neurosci Bull*, 39(2):328-342.  
<https://doi.org/10.1007/s12264-022-00960-4>
- Chevallier C, Kohls G, Troiani V, et al., 2012. The social motivation theory of autism. *Trends Cogn Sci*, 16(4):231-239.  
<https://doi.org/10.1016/j.tics.2012.02.007>
- Cho KIK, Shenton ME, Kubicki M, et al., 2016. Altered thalamo-cortical white matter connectivity: probabilistic tractography study in clinical-high risk for psychosis and first-episode psychosis. *Schizophr Bull*, 42(3):723-731.

- <https://doi.org/10.1093/schbul/sbv169>
- Chugani DC, Muzik O, Behen M, et al., 1999. Developmental changes in brain serotonin synthesis capacity in autistic and nonautistic children. *Ann Neurol*, 45(3):287-295. [https://doi.org/10.1002/1531-8249\(199903\)45:3<287::aid-ana3>3.0.co;2-9](https://doi.org/10.1002/1531-8249(199903)45:3<287::aid-ana3>3.0.co;2-9)
- Chung W, Choi SY, Lee E, et al., 2015. Social deficits in *IRSp53* mutant mice improved by NMDAR and mGluR5 suppression. *Nat Neurosci*, 18(3):435-443. <https://doi.org/10.1038/nn.3927>
- Chung Y, Haut KM, He G, et al., 2017. Ventricular enlargement and progressive reduction of cortical gray matter are linked in prodromal youth who develop psychosis. *Schizophr Res*, 189:169-174. <https://doi.org/10.1016/j.schres.2017.02.014>
- Courchesne E, Carper R, Akshoomoff N, 2003. Evidence of brain overgrowth in the first year of life in autism. *JAMA*, 290(3):337-344. <https://doi.org/10.1001/jama.290.3.337>
- Courchesne E, Campbell K, Solso S, 2011a. Brain growth across the life span in autism: age-specific changes in anatomical pathology. *Brain Res*, 1380:138-145. <https://doi.org/10.1016/j.brainres.2010.09.101>
- Courchesne E, Mouton PR, Calhoun ME, et al., 2011b. Neuron number and size in prefrontal cortex of children with autism. *JAMA*, 306(18):2001-2010. <https://doi.org/10.1001/jama.2011.1638>
- Curley AA, Arion D, Volk DW, et al., 2011. Cortical deficits of glutamic acid decarboxylase 67 expression in schizophrenia: clinical, protein, and cell type-specific features. *Am J Psychiatry*, 168(9):921-929. <https://doi.org/10.1176/appi.ajp.2011.11010052>
- David N, Schneider TR, Peiker I, et al., 2016. Variability of cortical oscillation patterns: a possible endophenotype in autism spectrum disorders? *Neurosci Biobehav Rev*, 71:590-600. <https://doi.org/10.1016/j.neubiorev.2016.09.031>
- de León Reyes NS, Sierra Díaz P, Nogueira R, et al., 2023. Corticotropin-releasing hormone signaling from prefrontal cortex to lateral septum suppresses interaction with familiar mice. *Cell*, 186(19):4152-4171.e31. <https://doi.org/10.1016/j.cell.2023.08.010>
- Diamond A, 2013. Executive functions. *Annu Rev Psychol*, 64:135-168. <https://doi.org/10.1146/annurev-psych-113011-143750>
- Donaldson ZR, Young LJ, 2008. Oxytocin, vasopressin, and the neurogenetics of sociality. *Science*, 322(5903):900-904. <https://doi.org/10.1126/science.1158668>
- Duffney LJ, Zhong P, Wei J, et al., 2015. Autism-like deficits in *Shank3*-deficient mice are rescued by targeting actin regulators. *Cell Rep*, 11(9):1400-1413. <https://doi.org/10.1016/j.celrep.2015.04.064>
- Ebstein RP, Israel S, Chew SH, et al., 2010. Genetics of human social behavior. *Neuron*, 65(6):831-844. <https://doi.org/10.1016/j.neuron.2010.02.020>
- Edden RAE, Muthukumaraswamy SD, Freeman TCA, et al., 2009. Orientation discrimination performance is predicted by GABA concentration and gamma oscillation frequency in human primary visual cortex. *J Neurosci*, 29(50):15721-15726. <https://doi.org/10.1523/JNEUROSCI.4426-09.2009>
- Eisenberg N, Fabes RA, Murphy B, et al., 1995. The role of emotionality and regulation in children's social functioning: a longitudinal study. *Child Dev*, 66(5):1360-1384. <https://doi.org/10.2307/1131652>
- Eisenberg N, Guthrie IK, Murphy BC, et al., 1999. Consistency and development of prosocial dispositions: a longitudinal study. *Child Dev*, 70(6):1360-1372. <https://doi.org/10.1111/1467-8624.00100>
- Eisenberg N, Fabes RA, Guthrie IK, et al., 2000. Dispositional emotionality and regulation: their role in predicting quality of social functioning. *J Pers Soc Psychol*, 78(1):136-157. <https://doi.org/10.1037/0022-3514.78.1.136>
- Fan ZX, Chang JR, Liang YL, et al., 2023. Neural mechanism underlying depressive-like state associated with social status loss. *Cell*, 186(3):560-576.e17. <https://doi.org/10.1016/j.cell.2022.12.033>
- Feinberg I, 1990. Cortical pruning and the development of schizophrenia. *Schizophr Bull*, 16(4):567-570. <https://doi.org/10.1093/schbul/16.4.567>
- Felix-Ortiz AC, Burgos-Robles A, Bhagat ND, et al., 2016. Bidirectional modulation of anxiety-related and social behaviors by amygdala projections to the medial prefrontal cortex. *Neuroscience*, 321:197-209. <https://doi.org/10.1016/j.neuroscience.2015.07.041>
- Filice F, Vörckel KJ, Sungur AÖ, et al., 2016. Reduction in parvalbumin expression not loss of the parvalbumin-expressing GABA interneuron subpopulation in genetic parvalbumin and shank mouse models of autism. *Mol Brain*, 9:10. <https://doi.org/10.1186/s13041-016-0192-8>
- Forbes CE, Grafman J, 2010. The role of the human prefrontal cortex in social cognition and moral judgment. *Annu Rev Neurosci*, 33:299-324. <https://doi.org/10.1146/annurev-neuro-060909-153230>
- Fornito A, Yücel M, Dean B, et al., 2009. Anatomical abnormalities of the anterior cingulate cortex in schizophrenia: bridging the gap between neuroimaging and neuropathology. *Schizophr Bull*, 35(5):973-993. <https://doi.org/10.1093/schbul/sbn025>
- Fries P, 2015. Rhythms for cognition: communication through coherence. *Neuron*, 88(1):220-235. <https://doi.org/10.1016/j.neuron.2015.09.034>
- Fujisawa S, Amarasingham A, Harrison MT, et al., 2008. Behavior-dependent short-term assembly dynamics in the medial prefrontal cortex. *Nat Neurosci*, 11(7):823-833. <https://doi.org/10.1038/nn.2134>
- Fung SJ, Webster MJ, Sivagnanasundaram S, et al., 2010. Expression of interneuron markers in the dorsolateral prefrontal cortex of the developing human and in schizophrenia. *Am J Psychiatry*, 167(12):1479-1488. <https://doi.org/10.1176/appi.ajp.2010.09060784>
- Gandal MJ, Edgar JC, Klook K, et al., 2012. Gamma synchrony: towards a translational biomarker for the treatment-resistant symptoms of schizophrenia. *Neuropharmacology*, 62(3):1504-1518. <https://doi.org/10.1016/j.neuropharm.2011.02.007>

- Gangopadhyay P, Chawla M, Dal Monte O, et al., 2021. Prefrontal-amygdala circuits in social decision-making. *Nat Neurosci*, 24(1):5-18.  
<https://doi.org/10.1038/s41593-020-00738-9>
- Geckeler KC, Barch DM, Karcher NR, 2022. Associations between social behaviors and experiences with neural correlates of implicit emotion regulation in middle childhood. *Neuropsychopharmacology*, 47(6):1169-1179.  
<https://doi.org/10.1038/s41386-022-01286-5>
- Giraldo-Chica M, Rogers BP, Damon SM, et al., 2018. Prefrontal-thalamic anatomical connectivity and executive cognitive function in schizophrenia. *Biol Psychiatry*, 83(6):509-517.  
<https://doi.org/10.1016/j.biopsych.2017.09.022>
- Gogtay N, Giedd JN, Lusk L, et al., 2004. Dynamic mapping of human cortical development during childhood through early adulthood. *Proc Natl Acad Sci USA*, 101(21):8174-8179.  
<https://doi.org/10.1073/pnas.0402680101>
- Goldman-Rakic PS, Selemon LD, 1997. Functional and anatomical aspects of prefrontal pathology in schizophrenia. *Schizophr Bull*, 23(3):437-458.  
<https://doi.org/10.1093/schbul/23.3.437>
- Han S, Tai C, Westenbroek RE, et al., 2012. Autistic-like behaviour in *Scn1a*<sup>+/-</sup> mice and rescue by enhanced GABA-mediated neurotransmission. *Nature*, 489(7416):385-390.  
<https://doi.org/10.1038/nature11356>
- Hanganu-Opatz IL, Klausberger T, Sigurdsson T, et al., 2023. Resolving the prefrontal mechanisms of adaptive cognitive behaviors: a cross-species perspective. *Neuron*, 111(7):1020-1036.  
<https://doi.org/10.1016/j.neuron.2023.03.017>
- Hashimoto T, Volk DW, Eggan SM, et al., 2003. Gene expression deficits in a subclass of GABA neurons in the prefrontal cortex of subjects with schizophrenia. *J Neurosci*, 23(15):6315-6326.  
<https://doi.org/10.1523/JNEUROSCI.23-15-06315.2003>
- Hashimoto T, Arion D, Unger T, et al., 2008. Alterations in GABA-related transcriptome in the dorsolateral prefrontal cortex of subjects with schizophrenia. *Mol Psychiatry*, 13(2):147-161.  
<https://doi.org/10.1038/sj.mp.4002011>
- Hazlett HC, Poe M, Gerig G, et al., 2005. Magnetic resonance imaging and head circumference study of brain size in autism: birth through age 2 years. *Arch Gen Psychiatry*, 62(12):1366-1376.  
<https://doi.org/10.1001/archpsyc.62.12.1366>
- Herbert MR, Ziegler DA, Deutsch CK, et al., 2003. Dissociations of cerebral cortex, subcortical and cerebral white matter volumes in autistic boys. *Brain*, 126(Pt 5):1182-1192.  
<https://doi.org/10.1093/brain/awg110>
- Huang AS, Rogers BP, Sheffield JM, et al., 2021. Characterizing effects of age, sex and psychosis symptoms on thalamocortical functional connectivity in youth. *NeuroImage*, 243:118562.  
<https://doi.org/10.1016/j.neuroimage.2021.118562>
- Huang WC, Zucca A, Levy J, et al., 2020. Social behavior is modulated by valence-encoding mPFC-amygdala sub-circuitry. *Cell Rep*, 32(2):107899.  
<https://doi.org/10.1016/j.celrep.2020.107899>
- Hughes JR, 2007. Autism: the first firm finding = underconnectivity? *Epilepsy Behav*, 11(1):20-24.  
<https://doi.org/10.1016/j.yebeh.2007.03.010>
- Hughes JR, 2008. A review of recent reports on autism: 1000 studies published in 2007. *Epilepsy Behav*, 13(3):425-437.  
<https://doi.org/10.1016/j.yebeh.2008.06.015>
- Kaczmarek LK, Zhang YL, 2017. Kv3 channels: enablers of rapid firing, neurotransmitter release, and neuronal endurance. *Physiol Rev*, 97(4):1431-1468.  
<https://doi.org/10.1152/physrev.00002.2017>
- Karlsgodt KH, Sanz J, van Erp TGM, et al., 2009. Re-evaluating dorsolateral prefrontal cortex activation during working memory in schizophrenia. *Schizophr Res*, 108(1-3):143-150.  
<https://doi.org/10.1016/j.schres.2008.12.025>
- Kehrer C, Maziashvili N, Dugladze T, et al., 2008. Altered excitatory-inhibitory balance in the NMDA-hypofunction model of schizophrenia. *Front Mol Neurosci*, 1:6.  
<https://doi.org/10.3389/neuro.02.006.2008>
- Kietzman HW, Gourley SL, 2023. How social information impacts action in rodents and humans: the role of the prefrontal cortex and its connections. *Neurosci Biobehav Rev*, 147:105075.  
<https://doi.org/10.1016/j.neubiorev.2023.105075>
- Kietzman HW, Trinoskey-Rice G, Blumenthal SA, et al., 2022. Social incentivization of instrumental choice in mice requires amygdala-prelimbic cortex-nucleus accumbens connectivity. *Nat Commun*, 13:4768.  
<https://doi.org/10.1038/s41467-022-32388-9>
- Kimoto S, Bazmi HH, Lewis DA, 2014. Lower expression of glutamic acid decarboxylase 67 in the prefrontal cortex in schizophrenia: contribution of altered regulation by Zif268. *Am J Psychiatry*, 171(9):969-978.  
<https://doi.org/10.1176/appi.ajp.2014.14010004>
- Kleinmans NM, Reiter MA, Neuhaus E, et al., 2016. Subregional differences in intrinsic amygdala hyperconnectivity and hypoconnectivity in autism spectrum disorder. *Autism Res*, 9(7):760-772.  
<https://doi.org/10.1002/aur.1589>
- Klune CB, Jin B, DeNardo LA, 2021. Linking mPFC circuit maturation to the developmental regulation of emotional memory and cognitive flexibility. *eLife*, 10:e64567.  
<https://doi.org/10.7554/eLife.64567>
- Kolb B, Mychasiuk R, Muhammad A, et al., 2012. Experience and the developing prefrontal cortex. *Proc Natl Acad Sci USA*, 109(S2):17186-17193.  
<https://doi.org/10.1073/pnas.1121251109>
- Kruse AO, Bustillo JR, 2022. Glutamatergic dysfunction in Schizophrenia. *Transl Psychiatry*, 12:500.  
<https://doi.org/10.1038/s41398-022-02253-w>
- Kumar A, Sundaram SK, Sivaswamy L, et al., 2010. Alterations in frontal lobe tracts and corpus callosum in young children with autism spectrum disorder. *Cereb Cortex*, 20(9):2103-2113.  
<https://doi.org/10.1093/cercor/bhp278>
- Lee E, Rhim I, Lee JW, et al., 2016. Enhanced neuronal activity in the medial prefrontal cortex during social approach behavior. *J Neurosci*, 36(26):6926-6936.



- <https://doi.org/10.1523/JNEUROSCI.0307-16.2016>
- Lee J, Chung C, Ha S, et al., 2015. *Shank3*-mutant mice lacking exon 9 show altered excitation/inhibition balance, enhanced rearing, and spatial memory deficit. *Front Cell Neurosci*, 9:94.  
<https://doi.org/10.3389/fncel.2015.00094>
- Lewis DA, 1997. Development of the prefrontal cortex during adolescence: insights into vulnerable neural circuits in schizophrenia. *Neuropsychopharmacology*, 16(6):385-398.  
[https://doi.org/10.1016/S0893-133X\(96\)00277-1](https://doi.org/10.1016/S0893-133X(96)00277-1)
- Li K, Nakajima M, Ibañez-Tallon I, et al., 2016. A cortical circuit for sexually dimorphic oxytocin-dependent anxiety behaviors. *Cell*, 167(1):60-72.e11.  
<https://doi.org/10.1016/j.cell.2016.08.067>
- Li Q, Takeuchi Y, Wang JL, et al., 2023. Reinstating olfactory bulb-derived limbic gamma oscillations alleviates depression-like behavioral deficits in rodents. *Neuron*, 111(13):2065-2075.e5.  
<https://doi.org/10.1016/j.neuron.2023.04.013>
- Liang B, Zhang LF, Barbera G, et al., 2018. Distinct and dynamic ON and OFF neural ensembles in the prefrontal cortex code social exploration. *Neuron*, 100(3):700-714.e9.  
<https://doi.org/10.1016/j.neuron.2018.08.043>
- Liang J, Xu W, Hsu YT, et al., 2015. Conditional neuroligin-2 knockout in adult medial prefrontal cortex links chronic changes in synaptic inhibition to cognitive impairments. *Mol Psychiatry*, 20(7):850-859.  
<https://doi.org/10.1038/mp.2015.31>
- Liu D, Tang QQ, Yin C, et al., 2018. Brain-derived neurotrophic factor-mediated projection-specific regulation of depressive-like and nociceptive behaviors in the mesolimbic reward circuitry. *Pain*, 159(1):175.  
<https://doi.org/10.1097/j.pain.0000000000001083>
- Liu L, Xu HF, Wang J, et al., 2020. Cell type-differential modulation of prefrontal cortical GABAergic interneurons on low gamma rhythm and social interaction. *Sci Adv*, 6(30):eaay4073.  
<https://doi.org/10.1126/sciadv.aay4073>
- Luna B, Minshew NJ, Garver KE, et al., 2002. Neocortical system abnormalities in autism: an fMRI study of spatial working memory. *Neurology*, 59(6):834-840.  
<https://doi.org/10.1212/wnl.59.6.834>
- Martínez K, Martínez-García M, Marcos-Vidal L, et al., 2020. Sensory-to-cognitive systems integration is associated with clinical severity in autism spectrum disorder. *J Am Acad Child Adolesc Psychiatry*, 59(3):422-433.  
<https://doi.org/10.1016/j.jaac.2019.05.033>
- Martínez-Losa M, Tracy TE, Ma KR, et al., 2018. Nav1.1-overexpressing interneuron transplants restore brain rhythms and cognition in a mouse model of Alzheimer's disease. *Neuron*, 98(1):75-89.e5.  
<https://doi.org/10.1016/j.neuron.2018.02.029>
- Martínez-Sanchis S, 2014. Neurobiological foundations of multisensory integration in people with autism spectrum disorders: the role of the medial prefrontal cortex. *Front Hum Neurosci*, 8:970.  
<https://doi.org/10.3389/fnhum.2014.00970>
- McGraw LA, Young LJ, 2010. The prairie vole: an emerging model organism for understanding the social brain. *Trends Neurosci*, 33(2):103-109.  
<https://doi.org/10.1016/j.tins.2009.11.006>
- McRae K, Gross JJ, 2020. Emotion regulation. *Emotion*, 20(1):1-9.  
<https://doi.org/10.1037/emo0000703>
- Michelsen KA, Prickaerts J, Steinbusch HWM, 2008. The dorsal raphe nucleus and serotonin: implications for neuroplasticity linked to major depression and Alzheimer's disease. *Prog Brain Res*, 172:233-264.  
[https://doi.org/10.1016/S0079-6123\(08\)00912-6](https://doi.org/10.1016/S0079-6123(08)00912-6)
- Millan MJ, Bales KL, 2013. Towards improved animal models for evaluating social cognition and its disruption in schizophrenia: the CNTRICS initiative. *Neurosci Biobehav Rev*, 37(9):2166-2180.  
<https://doi.org/10.1016/j.neubiorev.2013.09.012>
- Mitchell JP, Macrae CN, Banaji MR, 2006. Dissociable medial prefrontal contributions to judgments of similar and dissimilar others. *Neuron*, 50(4):655-663.  
<https://doi.org/10.1016/j.neuron.2006.03.040>
- Mitelman SA, Byne W, Kemether EM, et al., 2005. Metabolic disconnection between the mediodorsal nucleus of the thalamus and cortical Brodmann's areas of the left hemisphere in schizophrenia. *Am J Psychiatry*, 162(9):1733-1735.  
<https://doi.org/10.1176/appi.ajp.162.9.1733>
- Morris HM, Hashimoto T, Lewis DA, 2008. Alterations in somatostatin mRNA expression in the dorsolateral prefrontal cortex of subjects with schizophrenia or schizoaffective disorder. *Cereb Cortex*, 18(7):1575-1587.  
<https://doi.org/10.1093/cercor/bhm186>
- Moy SS, Nadler JJ, Perez A, et al., 2004. Sociability and preference for social novelty in five inbred strains: an approach to assess autistic-like behavior in mice. *Genes Brain Behav*, 3(5):287-302.  
<https://doi.org/10.1111/j.1601-1848.2004.00076.x>
- Müller NG, Machado L, Knight RT, 2002. Contributions of subregions of the prefrontal cortex to working memory: evidence from brain lesions in humans. *J Cogn Neurosci*, 14(5):673-686.  
<https://doi.org/10.1162/08989290260138582>
- Murray AJ, Woloszynowska-Fraser MU, Ansel-Bollepalli L, et al., 2015. Parvalbumin-positive interneurons of the prefrontal cortex support working memory and cognitive flexibility. *Sci Rep*, 5:16778.  
<https://doi.org/10.1038/srep16778>
- Murugan M, Jang HJ, Park M, et al., 2017. Combined social and spatial coding in a descending projection from the prefrontal cortex. *Cell*, 171(7):1663-1677.e16.  
<https://doi.org/10.1016/j.cell.2017.11.002>
- Nakajima M, Görlich A, Heintz N, 2014. Oxytocin modulates female sociosexual behavior through a specific class of prefrontal cortical interneurons. *Cell*, 159(2):295-305.  
<https://doi.org/10.1016/j.cell.2014.09.020>
- Nakazawa K, Jeevakumar V, Nakao K, 2017. Spatial and temporal boundaries of NMDA receptor hypofunction leading to schizophrenia. *npj Schizophr*, 3:7.  
<https://doi.org/10.1038/s41537-016-0003-3>

- Ogiwara I, Miyamoto H, Morita N, et al., 2007. Na<sub>v</sub>1.1 localizes to axons of parvalbumin-positive inhibitory interneurons: a circuit basis for epileptic seizures in mice carrying an *Scn1a* gene mutation. *J Neurosci*, 27(22):5903-5914. <https://doi.org/10.1523/JNEUROSCI.5270-06.2007>
- Ohnishi T, Matsuda H, Hashimoto T, et al., 2000. Abnormal regional cerebral blood flow in childhood autism. *Brain*, 123(Pt 9):1838-1844. <https://doi.org/10.1093/brain/123.9.1838>
- Okuyama T, Kitamura T, Roy DS, et al., 2016. Ventral CA1 neurons store social memory. *Science*, 353(6307):1536-1541. <https://doi.org/10.1126/science.aaf7003>
- Padilla-Coreano N, Batra K, Patarino M, et al., 2022. Cortical ensembles orchestrate social competition through hypothalamic outputs. *Nature*, 603(7902):667-671. <https://doi.org/10.1038/s41586-022-04507-5>
- Palagi E, Burghardt GM, Smuts B, et al., 2016. Rough-and-tumble play as a window on animal communication. *Biol Rev*, 91(2):311-327. <https://doi.org/10.1111/brv.12172>
- Pellis SM, Pellis VC, Ham JR, et al., 2023. Play fighting and the development of the social brain: the rat's tale. *Neurosci Biobehav Rev*, 145:105037. <https://doi.org/10.1016/j.neubiorev.2023.105037>
- Rafice F, Rezvani Habibabadi R, Motaghi M, et al., 2022. Brain MRI in autism spectrum disorder: narrative review and recent advances. *J Magn Reson Imaging*, 55(6):1613-1624. <https://doi.org/10.1002/jmri.27949>
- Rane P, Cochran D, Hodge SM, et al., 2015. Connectivity in autism: a review of MRI connectivity studies. *Harv Rev Psychiatry*, 23(4):223-244. <https://doi.org/10.1097/HRP.0000000000000072>
- Rein B, Tan T, Yang FW, et al., 2021. Reversal of synaptic and behavioral deficits in a 16p11.2 duplication mouse model via restoration of the GABA synapse regulator *Npas4*. *Mol Psychiatry*, 26(6):1967-1979. <https://doi.org/10.1038/s41380-020-0693-9>
- Rudebeck PH, Walton ME, Millette BHP, et al., 2007. Distinct contributions of frontal areas to emotion and social behaviour in the rat. *Eur J Neurosci*, 26(8):2315-2326. <https://doi.org/10.1111/j.1460-9568.2007.05844.x>
- Sacco R, Gabriele S, Persico AM, 2015. Head circumference and brain size in autism spectrum disorder: a systematic review and meta-analysis. *Psychiatry Res Neuroimaging*, 234(2):239-251. <https://doi.org/10.1016/j.psychres.2015.08.016>
- Sarawagi A, Soni ND, Patel AB, 2021. Glutamate and GABA homeostasis and neurometabolism in major depressive disorder. *Front Psychiatry*, 12:637863. <https://doi.org/10.3389/fpsy.2021.637863>
- Sceniak MP, Lang M, Enomoto AC, et al., 2016. Mechanisms of functional hypoconnectivity in the medial prefrontal cortex of *Mecp2* null mice. *Cereb Cortex*, 26(5):1938-1956. <https://doi.org/10.1093/cercor/bhv002>
- Scheggia D, Managò F, Maltese F, et al., 2020. Somatostatin interneurons in the prefrontal cortex control affective state discrimination in mice. *Nat Neurosci*, 23(1):47-60. <https://doi.org/10.1038/s41593-019-0551-8>
- Selemon LD, Zecevic N, 2015. Schizophrenia: a tale of two critical periods for prefrontal cortical development. *Transl Psychiatry*, 5(8):e623. <https://doi.org/10.1038/tp.2015.115>
- Selimbeyoglu A, Kim CK, Inoue M, et al., 2017. Modulation of prefrontal cortex excitation/inhibition balance rescues social behavior in *CNTNAP2*-deficient mice. *Sci Transl Med*, 9(401):eaa6733. <https://doi.org/10.1126/scitranslmed.aah6733>
- Shemesh Y, Sztainberg Y, Forkosh O, et al., 2013. High-order social interactions in groups of mice. *eLife*, 2:e00759. <https://doi.org/10.7554/eLife.00759>
- Sohal VS, Zhang F, Yizhar O, et al., 2009. Parvalbumin neurons and gamma rhythms enhance cortical circuit performance. *Nature*, 459(7247):698-702. <https://doi.org/10.1038/nature07991>
- Su MH, Hu KJ, Liu W, et al., 2024. Theta oscillations support prefrontal-hippocampal interactions in sequential working memory. *Neurosci Bull*, 40:147-156. <https://doi.org/10.1007/s12264-023-01134-6>
- Sun QT, Li XN, Li AN, et al., 2020. Ventral hippocampal-prefrontal interaction affects social behavior via parvalbumin positive neurons in the medial prefrontal cortex. *iScience*, 23(3):100894. <https://doi.org/10.1016/j.isci.2020.100894>
- Tan YL, Singhal SM, Harden SW, et al., 2019. Oxytocin receptors are expressed by glutamatergic prefrontal cortical neurons that selectively modulate social recognition. *J Neurosci*, 39(17):3249-3263. <https://doi.org/10.1523/JNEUROSCI.2944-18.2019>
- Uddin LQ, Supekar K, Menon V, 2013. Reconceptualizing functional brain connectivity in autism from a developmental perspective. *Front Hum Neurosci*, 7:458. <https://doi.org/10.3389/fnhum.2013.00458>
- Varlinkaya EI, Spear LP, Spear NE, 1999. Social behavior and social motivation in adolescent rats: role of housing conditions and partner's activity. *Physiol Behav*, 67(4):475-482. [https://doi.org/10.1016/s0031-9384\(98\)00285-6](https://doi.org/10.1016/s0031-9384(98)00285-6)
- Vawter MP, Crook JM, Hyde TM, et al., 2002. Microarray analysis of gene expression in the prefrontal cortex in schizophrenia: a preliminary study. *Schizophr Res*, 58(1):11-20. [https://doi.org/10.1016/s0920-9964\(01\)00377-2](https://doi.org/10.1016/s0920-9964(01)00377-2)
- Vissers ME, Cohen MX, Geurts HM, 2012. Brain connectivity and high functioning autism: a promising path of research that needs refined models, methodological convergence, and stronger behavioral links. *Neurosci Biobehav Rev*, 36(1):604-625. <https://doi.org/10.1016/j.neubiorev.2011.09.003>
- Wang F, Zhu J, Zhu H, et al., 2011. Bidirectional control of social hierarchy by synaptic efficacy in medial prefrontal cortex. *Science*, 334(6056):693-697. <https://doi.org/10.1126/science.1209951>
- Wang F, Kessels HW, Hu HL, 2014. The mouse that roared: neural mechanisms of social hierarchy. *Trends Neurosci*, 37(11):674-682.

- <https://doi.org/10.1016/j.tins.2014.07.005>
- Wang W, Rein B, Zhang F, et al., 2018. Chemogenetic activation of prefrontal cortex rescues synaptic and behavioral deficits in a mouse model of 16p11.2 deletion syndrome. *J Neurosci*, 38(26):5939-5948.  
<https://doi.org/10.1523/JNEUROSCI.0149-18.2018>
- Weinberger DR, Berman KF, 1996. Prefrontal function in schizophrenia: confounds and controversies. *Philos Trans Roy Soc B Biol Sci*, 351(1346):1495-1503.  
<https://doi.org/10.1098/rstb.1996.0135>
- Woo TUW, Kim AM, Viscidi E, 2008. Disease-specific alterations in glutamatergic neurotransmission on inhibitory interneurons in the prefrontal cortex in schizophrenia. *Brain Res*, 1218:267-277.  
<https://doi.org/10.1016/j.brainres.2008.03.092>
- Woodward ND, Heckers S, 2016. Mapping thalamocortical functional connectivity in chronic and early stages of psychotic disorders. *Biol Psychiatry*, 79(12):1016-1025.  
<https://doi.org/10.1016/j.biopsych.2015.06.026>
- Woodward ND, Karbasforoushan H, Heckers S, 2012. Thalamocortical dysconnectivity in schizophrenia. *Am J Psychiatry*, 169(10):1092-1099.  
<https://doi.org/10.1176/appi.ajp.2012.12010056>
- Yizhar O, Fenno LE, Prigge M, et al., 2011. Neocortical excitation/inhibition balance in information processing and social dysfunction. *Nature*, 477(7363):171-178.  
<https://doi.org/10.1038/nature10360>
- Zhang CY, Zhu H, Ni ZY, et al., 2022. Dynamics of a disinhibitory prefrontal microcircuit in controlling social competition. *Neuron*, 110(3):516-531.e6.  
<https://doi.org/10.1016/j.neuron.2021.10.034>
- Zhang MQ, Palaniyappan L, Deng MJ, et al., 2021. Abnormal thalamocortical circuit in adolescents with early-onset schizophrenia. *J Am Acad Child Adolesc Psychiatry*, 60(4):479-489.  
<https://doi.org/10.1016/j.jaac.2020.07.903>
- Zhou TT, Sandi C, Hu HL, 2018. Advances in understanding neural mechanisms of social dominance. *Curr Opin Neurobiol*, 49:99-107.  
<https://doi.org/10.1016/j.conb.2018.01.006>